



Cardiovascular abnormalities in primary immunodeficiency diseases

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ABSTRACT

In recent years, increasing numbers of patients with primary immune deficiency (PID) are being recognized as also suffering from cardiovascular system (CVS) abnormalities. These CVS defects might be explained by infectious or autoimmune etiologies, as well as by the role of specific genes and the immune system in the development and function of CVS tissues. Here, we provide the first comprehensive review of the clinical, potentially pathogenic mechanisms, and the management of PID, as well as the associated immune and CVS defects. In addition to some well-known associations of PID with CVS abnormalities, such as DiGeorge syndrome and CHARGE anomaly, we describe the cardiac defects associated with Omenn syndrome, calcium channel deficiencies, DNA repair defects, common variable immunodeficiency, Roifman syndrome, various neutrophil/macrophage defects, FADD deficiency, and HOIL1 deficiency. Moreover, we detail the vascular abnormalities recognized in chronic mucocutaneous candidiasis, chronic granulomatous disease, Wiskott–Aldrich syndrome, Schimke immuno-osseous dysplasia, hyper-IgE syndrome, MonoMAC syndrome, and X-linked lymphoproliferative disease. In conclusion, the expanding spectrum of PID requires increased alertness to the possibility of CVS involvement as an important contributor to the diagnosis and management of these patients.

Introduction

Primary immunodeficiency diseases (PID) encompass a broad spectrum of inheritable disorders that are commonly associated with increased susceptibility to infections, immune dysregulation, and malignancies. Early recognition and correction of the immune deficiency is crucial. Patients with profound T-cell immune deficiency such as severe combined immunodeficiency (SCID) should be treated by allogeneic hematopoietic stem cell transplantation (HSCT) or autologous gene therapy, whereas antibody deficiency can be corrected by immunoglobulin replacement.

The important role of the immune system in the development and function of many organs as well as the effects of gene defects in nonimmune tissues contributes to the multi-system abnormalities often observed in patients suffering from PID. Additionally, the nonimmune abnormalities can be a clue to establishing the specific etiology of the PID, anticipating potential complications or choosing best management options. Indeed, nonimmune neurological, bone, gastrointestinal, skin, and endocrine abnormalities are often associated with various PID, such as purine nucleoside phosphorylase deficiency, adenosine deaminase deficiency, cartilage–hair hypoplasia, ectodermal dysplasia,

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etc. In recent years, particularly with the significant increase in PID for which molecular defects are identified, many PID conditions are recognized to be associated with cardiovascular system (CVS) abnormalities.

Here we provide a comprehensive review of PID associated with CVS defects. The review was performed using the PubMed database. The International Union of Immunological Societies Expert Committee for Primary Immunodeficiency classification system (Bousfiha et al. 2013) was used to systematically search for articles reporting PID associated with cardiac or vascular malformations. Additional associations were identified through an open PubMed search by combining terms such as “primary immunodeficiency” and “cardiac” or “heart” or “vascular”. CVS complications secondary to infectious etiologies as well as disorders caused predominantly or typically by rheumatologic and autoimmune mechanisms were excluded.

The review is intended for both cardiologists and immunologists. Therefore, following a brief description of the clinical features and pathogenesis of the specific condition, the immunological and CVS abnormalities are provided as well as management strategies.

Also, as a useful reference, we included 3 Appendices that summarize the cardiac and vascular abnormalities associated with PID (Appendix A1), the PID associated with cardiac abnormalities (Appendix A2), and the PID associated with vascular abnormalities (Appendix A3). Additionally, Appendix B1 summarizes the acronyms used in this review.

Predominantly T lineage defects

Omenn syndrome

Description: Omenn syndrome (OMIM# 603554), a combination of generalized severe erythroderma, lymphadenopathy, eosinophilia, and profound immunodeficiency in infancy, was first described in 1965. Other manifestations include hepatosplenomegaly, failure to thrive, diarrhea, and alopecia. Laboratory investigations often reveal eosinophilia and elevated IgE (Villa et al. 2008).

Pathogenesis: Omenn syndrome can occur with practically all profound T-cell immune deficiencies, such as those caused by defects in recombinant activating genes (*RAG*) 1 and 2, *ARTEMIS* (*DCLRE1C* gene), the RNA component of mitochondrial RNA processing

endonuclease gene (*RMRP*), *DNA Ligase IV*, interleukin 7 receptor α , common γ chain, CD3 δ , adenosine deaminase, and purine nucleoside phosphorylase deficiencies, DiGeorge syndrome (DGS), etc. (Villa et al. 1998, 2008; Ege et al. 2005; Zhang et al. 2005; Grunebaum et al. 2008). Common to these conditions is the ability of some T cells to mature in the thymus, escape proper selection, and expand in the periphery (Aleman et al. 2001; Poliani et al. 2009).

Immune abnormalities: Patients with Omenn syndrome have severely impaired cellular and humoral immunity; however, in contrast to typical severe combined immunodeficiency, T-cell numbers may be normal or increased. Proliferation of T cells ex vivo in response to various stimuli is often depressed, but might be present. T-cell receptor excision circles (TREC), which represent new thymus emigrants, are markedly reduced (Roifman et al. 2012). Significantly limited T-cell diversity and T-cell clonal expansion, increased IgE, and eosinophils are additional hallmarks of Omenn syndrome (Somech et al. 2009).

CVS abnormalities: Cardiac manifestations of Omenn syndrome have rarely been described. An infant with Omenn syndrome was found to have biventricular hypertrophy, impaired left ventricular systolic function, and severe sinus bradycardia, possibly secondary to endomyocardial disease caused by eosinophilia (Bruckmann et al. 1991).

Treatment: Antimicrobial and anti-inflammatory medications such as systemic steroids or cyclosporine A together with strict isolation can provide temporary benefit. Ultimately, Omenn syndrome can only be cured by HSCT, preferably from a human leucocyte antigen (HLA)-matched sibling or unrelated donor (Nahum et al. 2009).

Calcium channel deficiency

Description: ORAI-1 deficiency (OMIM# 610277) is an autosomal recessive form of SCID, characterized by recurrent infections in infancy, congenital muscular hypotonia, and anhydrotic ectodermal dysplasia (Feske et al. 2010). It is also associated with autoimmunity, developmental delay, and failure to thrive (McCarl et al. 2009).

Pathogenesis: This clinical syndrome is caused by a mutation in the *ORAI1* gene, encoding part of the calcium release activated calcium channel. T cells rely on

this channel for calcium influx for subsequent activation. Thus, mutations in *ORAI1* lead to severely impaired T-cell function. Calcium influx secondary to calcium release activated calcium channel function is also crucial for skeletal muscle function and (or) differentiation, leading to the congenital myopathy (McCarl et al. 2009; Feske et al. 2010). Ectodermal dysplasia is characterized by an inability to sweat and impaired thermoregulation, for which the precise mechanism is not clear.

Immune abnormalities: ORAI-1 deficient patients suffer from recurrent viral (particularly herpes viruses), bacterial, and fungal infections despite having normal T-cell and B-cell numbers. Lymphocyte development is unaffected; however, T-cell activation as well as the functions of other immune cells, including B cells, are severely impaired. Natural killer (NK) cell functions, including degranulation, cytokine production, and cytolytic activity may also be impaired (Notarangelo 2013).

CVS abnormalities: CVS abnormalities have not been reported in ORAI-1 deficient patients; however, inactivation of *ORAI1* in Zebra fish causes reduced ventricular systolic function, bradycardia, and heart failure (Volkers et al. 2012), suggesting that cardiomyopathy may co-exist with the skeletal myopathy.

A similar, albeit less severe, immunodeficiency is present among patients with *STIM1* mutations (OMIM# 605921) that also disrupt calcium entry; however, cardiac defects are not reported (Feske et al. 2010).

Treatment: HSCT can correct the immunodeficiency; however, the skin and muscle abnormalities persist.

Chronic mucocutaneous candidiasis

Description: Chronic mucocutaneous candidiasis (CMCC) is a group of disorders characterized by recurrent *Candida* infections of the skin, nails, and mucous membranes, which may appear in infancy or later in life and are often accompanied by autoimmune endocrine abnormalities such as hypo-parathyroidism and adrenal insufficiency. Indeed, some patients are categorized as suffering from autoimmune polyendocrinopathy–candidiasis–ectodermal dystrophy or autoimmune polyendocrine syndrome type 1. Patients may also develop chronic keratitis, eczema, and other autoimmune phenomena (Liu et al. 2011).

Pathogenesis: CMCC phenotype is associated with mutations in several genes. An autosomal recessive variant of CMCC is caused by defects in the autoimmune regulator (*AIRE*) gene (OMIM# 240300) encoding a transcription factor expressed in the medulla of the thymus and is thought to be important for presentation of tissue-specific self-antigens to maturing thymocytes. *DECTIN-1* and *CARD9* deficiency have also been associated with autosomal recessive CMCC. Mutations in *STAT1*, a transcription factor involved in multiple signaling pathways, are the main cause of autosomal dominant CMCC (OMIM# 600555), although the precise mechanism for the susceptibility to candidiasis is still not clear (Liu et al. 2011).

Immune abnormalities: Immune dysfunction in CMCC is variable. Typically, *Candida* infections are limited to the mucocutaneous membranes and are not invasive, although patients may display T-cell deficiency, manifesting with increased susceptibility to cytomegalovirus and other viruses (Nahum et al. 2012) as well as disseminated coccidioidomycosis and histoplasmosis (Sampaio et al. 2013). Some patients do not produce appropriate antibodies, resulting in increased frequency of sinopulmonary infections. Laboratory findings in patients with CMCC include normal or reduced in vitro and in vivo T-cell responses to *Candida* antigens, elevated IgE, and detection of various autoantibodies (Kirkpatrick, 2001; Puel et al. 2010). In addition to glandular damage, there have been reports of autoimmune cytopenias, hepatitis, colitis, etc. (Hori et al. 2012). Some patients might also develop thymoma or hemophagocytic lymphohistiocytosis or present as Omenn syndrome (Cavadini et al. 2005).

CVS abnormalities: CMCC is associated with vasculitis, particularly cerebral vasculitis, with severe neurological complications (Grouhi et al. 1998; Nagashima et al. 2000). A brain biopsy in one patient demonstrated perivascular mononuclear cell infiltrates, which is suggestive of endarteritis (Grouhi et al. 1998). Subsequently, patients with CMCC with dominant *STAT1* mutations were noted to suffer from cerebral aneurysms and infarctions (Marazzi et al. 2008; Uzel et al. 2013). It is still not clear whether these cerebral abnormalities are caused by unrecognized infections, autoimmune phenomena, or structural defect directly related to either the gene defect or the potential involvement of other tissues.

Treatment: Oral and topical antifungal medications are the mainstay of treatment for CMCC (Kirkpatrick

2001). Immunoglobulin replacement has been beneficial for patients with antibody deficiency, whereas immune suppressive medications have been used for the autoimmune phenomena. The benefit of HSCT for the very severe forms of CMCC is still not established.

STK-4 deficiency

Description: Autosomal recessive mutations in *STK4*, also known as *MST1* (OMIM# 614868), were found among consanguineous Iranian and Turkish families with lymphopenia, intermittent neutropenia, and structural cardiac defects (Abdollahpour et al. 2012; Nehme et al. 2012).

Pathogenesis: Increased apoptosis of STK-4 deficient T cells and granulocytes may explain the reduced numbers of cells.

Immune abnormalities: Patients suffer from recurrent bacterial infections, including sinusitis, pneumonia, and meningitis, as well as herpes and papilloma virus infections, mucocutaneous candidiasis, cutaneous warts, and skin abscesses. Patients may present in infancy or childhood. Dermatitis and autoimmunity, including hemolytic anemia, also develop in some patients. Immune evaluations demonstrate lymphopenia with paucity of T cells, particularly CD4⁺ cells. Proliferation in response to stimulation with mitogens or antigens is difficult to assess because of the rapid death of STK-4 deficient T cells in vitro. B-cell numbers are reduced; however, IgG and IgE levels are increased and production of various antibodies is appropriate. The neutropenia might be intermittent in some patients. Contrary to many other forms of congenital neutropenia, the bone marrow of the patients with STK-4 deficiency appears normal. Epstein-Barr virus associated lymphadenopathy, phenotypically resembling lymphoplasmacytic lymphoma can also occur (Abdollahpour et al. 2012).

CVS abnormalities: Atrial septal defect type II, patent foramen ovale as well as mitral, tricuspid, and pulmonary insufficiency can be identified in some patients. Interestingly, STK-4 deficient mice develop dilated cardiomyopathy secondary to increased apoptosis of myocytes (Yamamoto et al. 2003).

Treatment: Antimicrobial and antiviral prophylaxis as well as immunoglobulin infusions may help prevent the frequent infections in STK-4 deficient patients. Close monitoring for lymphoproliferative disease and

cardiac defects are warranted. HSCT appears to cure the immunodeficiency (Nehme et al. 2012).

Lymphocyte-specific protein tyrosine kinase deficiency

A single patient with homozygous lymphocyte-specific protein tyrosine kinase (*LCK*) mutation (OMIM# 153390) who suffered from recurrent infections and retinal vasculitis was identified by Hauck et al. (2012).

Pathophysiology: Despite the important role of *LCK* in T cells, some T cells do develop and escape the thymus, possibly contributing to the immune dysregulation observed in *LCK* deficiency.

Immune abnormalities: The patient suffered from recurrent respiratory tract infections, protracted gut disease, failure to thrive, nodular skin lesions, arthritis, and autoimmune thrombocytopenia from an early age. Immune evaluations revealed severe CD4⁺ T-cell lymphopenia with oligoclonal TCRαβ⁺ T cells, increase in central memory CD4⁺ cell, and “exhausted” T-effector memory CD8⁺ cells. Activation of T cells with anti-CD3 demonstrated markedly reduced early tyrosine phosphorylation events and abrogated Ca²⁺ mobilization resulting in a severe proliferation defect. IgM levels were increased, whereas antibodies to vaccines were low. Autoantibodies to multiple self-antigens were present. The low number of Treg cells and the reduced activation-induced cell death of the patient’s T cells might have contributed to the immune dysregulation (Hauck et al. 2012).

CVS abnormalities: The patient suffered from pericarditis, possibly related to capillary leak syndrome as well as retinal vasculitis (Hauck et al. 2012).

Treatment: Antibiotic prophylaxis and immunoglobulin replacement are warranted. This single patient with *LCK* deficiency died shortly after HSCT from veno-occlusive disease.

DNA repair defects

Ataxia-telangiectasia

Description: Ataxia-telangiectasia (AT) is a well-described autosomal recessive disorder characterized by cerebellar ataxia, oculomotor apraxia, telangiectasia, and immunodeficiency (OMIM# 208900). Patients’ cells are also highly sensitive to irradiation. Progressive

ataxia is often noted at 2–3 years of age. Other neurologic features include nystagmus, dysarthria, and choreoathetosis. Telangiectasia in the eyes and skin typically follow the neurological symptoms. Patients are at significant risk for malignancy. Elevated α -feto-protein blood levels are very helpful in establishing the diagnosis of AT beyond the first few months of life, although other genetic defects such as ataxia with oculomotor apraxia type 1 and type 2 can also cause ataxia and increased α -fetoprotein (Nanetti et al. 2013).

Pathogenesis: AT is caused by defects in the AT mutated protein, a kinase with significant role in sensing and repairing double-stranded DNA breaks. Accordingly, AT mutated protein defects lead to abnormal DNA repair, resulting in accumulation of damaged genetic material in neuronal and other cells (Perlman et al. 2012).

Immune abnormalities: The immunodeficiency is variable. Sinopulmonary infections are frequent. Typical laboratory manifestations include decreased IgA, IgE, and IgG subclasses and antibody responses, whereas IgM might be elevated, resulting in a phenotype similar to that of hyper-IgM syndrome. Lymphopenia, depressed T-cell function, and reduced TREC can also be found, suggesting that some patients might be diagnosed through SCID newborn screening.

CVS abnormalities: Mitral valve prolapse, mitral regurgitation, and tricuspid regurgitation are found in few patients with AT (Bastianon et al. 1993). Interestingly, carriers of mutations in a single allele AT mutated kinase are more susceptible to ischemic heart disease, from an as yet unknown cause (Su and Swift 2000).

Treatment: Currently, there is no cure for AT. Patients should avoid exposure to ionizing radiation. Physical, occupational, and speech therapy, as well as a healthy diet and lifestyle can improve quality of life (Perlman et al. 2012). Antibiotic prophylaxis or immunoglobulin therapy can aid patients with recurrent infections or documented evidence of cellular and humoral immune deficiency.

Nijmegen breakage syndrome

Description: Nijmegen breakage syndrome (OMIM# 251260) is a rare autosomal recessive disorder characterized by chromosomal instability. Common clinical features include microcephaly, immunodeficiency,

radiation hypersensitivity, and predisposition to malignancy. Patients with Nijmegen breakage syndrome may also display typical bird-like facial features with a prominent mid-face, micrognathia, and beaked nose. Nijmegen breakage syndrome is also associated with growth delay and premature ovarian failure in females (Chrzanowska et al. 2012).

Pathogenesis: Nijmegen breakage syndrome is caused by mutations in the *NBS1* gene, encoding the Nibrin protein. Nibrin is a component of the MRN (MRE11–RAD50–NBN) complex that is critical to the DNA damage response system, along with the AT mutated protein. Thus, defective Nibrin leads to impaired DNA repair and a phenotype similar to AT (Chrzanowska et al. 2012).

Immune abnormalities: There are significant variations in the immunodeficiency among Nijmegen breakage syndrome patients. Most frequently, patients have reduced immunoglobulin levels. B and T cells may also be decreased (Chrzanowska et al. 2012).

CVS abnormalities: One patient with Nijmegen breakage syndrome was reported to suffer from a large sub-aortic ventricular septum defect (Tekin et al. 2002), possibly related to the abundant expression of *NBS1* in the cardiac muscle, as demonstrated in mouse embryos (Wilda et al. 2000).

Treatment: Antibiotic prophylaxis and immunoglobulin replacement therapy may prevent infections, while physiotherapy and occupational therapy may help improve quality of life. As in AT, avoiding exposure to ionizing radiation and regular screening for malignancy is very important (Chrzanowska et al. 2012). Recently, favorable outcome following HSCT was reported for few patients with Nijmegen breakage syndrome with severe immunodeficiency or malignancy (Albert et al. 2010).

Predominantly antibody deficiencies

Common variable immunodeficiency

Description: Common variable immunodeficiency (CVID) is a heterogeneous group of disorders characterized by hypogammaglobulinemia, abnormal production of antibodies, and no other known cause for the immune defect. Infections typically occur in early- to mid-adulthood but can present in childhood or even among aging individuals. Few CVIDs are secondary to

single gene defects that interfere with B-cell development or function, yet the majority of cases have no underlying cause (Cunningham-Rundles 2012).

Immune abnormalities: Most patients suffer from recurrent sinopulmonary infections, as well as arthritis, osteomyelitis, meningitis, or sepsis caused by bacterial and viral pathogens. Many patients come to medical attention due to inflammatory or autoimmune complications (Cunningham-Rundles 2012). The latter include organ-specific and hematological autoimmune cytopenias, lymphocyte infiltration into the lung and gastrointestinal tract, lymphadenopathy, splenomegaly, and granulomatous disease (Conley et al. 2009). Typically, IgG is reduced, whereas IgA and IgM may be normal, elevated, or reduced. It is important to exclude secondary causes for low IgG, such as gastrointestinal or urine loss, medication effects, malignancy, etc. Specific antibodies to infections or vaccinations are typically absent. Autoantibodies may appear with or without clinical autoimmune manifestations. The number of circulating B cells is usually normal (Conley et al. 2009) while kappa-deleting recombination excision circles are reduced in some patients (Kamae et al. 2013).

CVS abnormalities: CVID is not classically associated with CVS disease; however, few patients with CVID have aortic root dilatation (Johnston et al. 2004). A 16-year-old boy with CVID was reported to have an aortic aneurysm extending to the renal arteries (Kashef et al. 2011), whereas giant cell myocarditis, a rare aggressive form of autoimmune myocarditis, was found in a teenager with CVID (Laufs et al. 2002). Lastly, a 41-year-old woman, diagnosed with CVID later in life was found to have an asymptomatic right-sided aortic arch (Yalcin et al. 2004).

Treatment: The mainstay treatment for primary B-cell defects, including CVID, is immunoglobulin replacement. Patients should also receive prompt and appropriate antibiotic therapy for infections. Antibiotic prophylaxis against respiratory infections might also be beneficial (Hoernes et al. 2011).

TWEAK deficiency

Autosomal-dominant deficiency in tumor necrosis factor (TNF)-like weak inducer of apoptosis (TWEAK) can cause recurrent infections and impaired antibody responses to protein and polysaccharide vaccines (Wang et al. 2013). Patients may also develop chronic

thrombocytopenia, intermittent neutropenia, and multiple warts. Patients do not suffer from CVS abnormalities, yet TWEAK, a member of the TNF superfamily ligands, has a direct effect on cardiomyocytes (Jain et al. 2009).

Pathogenesis: The immune abnormalities are attributed to inhibition of B-cell activating factor signaling in B cells secondary to impaired activation of the NF- κ B and MAPK pathways. TWEAK action on the cardiac Fn14 receptor is thought to cause cardiomyocyte elongation and cardiac fibrosis.

Immune abnormalities: Patients with TWEAK deficiency suffer from recurrent sinopulmonary infections from infancy, as well as meningitis and osteomyelitis. The frequency of infections may decrease with age. Immune evaluations demonstrate low to normal IgG levels as well as low levels of IgM and IgA. Antibody responses to T-cell dependent and polysaccharide antigens are absent. Nearly all of the B cells are of naive phenotype.

CVS abnormalities: Elevated circulating TWEAK levels were found among patients with idiopathic dilated cardiomyopathy (Jain et al. 2009).

Treatment: Antibiotic and immunoglobulin treatment and prophylaxis may help the infections. Careful evaluation for the possibility of developing CVS abnormalities is needed.

Predominantly defects of neutrophils/macrophages number and function

Severe congenital neutropenia type 4/ G6PC3 deficiency

Description: Severe congenital neutropenia (SCN) type 4 (OMIM# 612541) is a rare autosomal recessive disorder characterized by neutropenia, recurrent bacterial infections, and increased risk of leukemia, structural heart defects, and urogenital malformations (Boztug et al. 2009). Prominent superficial venous pattern, myopathy, increased skin laxity, congenital hydronephrosis, and thrombocytopenia, which might be intermittent, are observed in some patients (Boztug et al. 2009; Banka et al. 2011).

Pathogenesis: SCN4 is caused by mutations in the glucose-6-phosphatase, catalytic subunit 3 (G6PC3)

gene, resulting in decreased function of the glucose-6-phosphatase enzyme and possibly decreased intracellular glucose concentrations (Boztug et al. 2009; Banka et al. 2011). *G6PC3* deficiency leads to increased neutrophil apoptosis. Bone marrow examination of some patients with *G6PC3* deficiency demonstrates maturation arrest of neutrophils, whereas in other patients myelokathexis is evident. Enhanced apoptosis may also be responsible for the congenital malformations associated with *SCN4* (Boztug et al. 2009; Banka et al. 2011).

Immune abnormalities: Neonatal sepsis, frequent sinopulmonary infections, oral ulcers, etc., are attributed to the severe neutropenia. One patient was also recently described with T-cell lymphopenia (Begin et al. 2013).

CVS abnormalities: *SCN4* is associated with structural heart abnormalities including atrial septum defects, patent ductus arteriosus, mitral regurgitation, anomalous pulmonary veins, pulmonary stenosis, and cor triatriatum (Boztug et al. 2009; Banka et al. 2011).

Treatment: Neutrophil counts often improve following granulocyte cell stimulating factor administration (McDermott et al. 2010). Cardiac and (or) urogenital malformations may need surgical intervention.

Barth syndrome

Description: Barth syndrome (OMIM# 302060) is an X-linked disorder characterized by neutropenia, cardiac disease, skeletal myopathy, developmental delay, and failure to thrive. Organic aciduria and hypocholesterolemia may also be present. The phenotype varies in severity among patients and may change over time (Schlame and Ren 2006; Jefferies 2013).

Pathogenesis: Barth syndrome is caused by a mutation in the *TAFAZZIN* gene on chromosome Xq28. This results in impaired phospholipid metabolism and disruption of mitochondrial biogenesis. Muscle and heart tissue have high-energy turnover and require strict structural organization of mitochondria. Thus, the abnormal mitochondria might lead to the skeletal and cardiac myopathy. Abnormal mitochondria also cause exposure of phosphatidylserine in cell membranes, leading to increased clearance of neutrophils by tissue macrophages (Schlame and Ren 2006; Finsterer and Frank 2013).

Immune abnormalities: Neutropenia is the main immunological feature. In some cases, it is intermittent or not present at all (Spencer et al. 2006).

CVS abnormalities: Dilated cardiomyopathy is the classic cardiac feature associated with Barth syndrome. Less common manifestations include hypertrophic cardiomyopathy, left ventricular noncompaction, endocardial fibroelastosis, ventricular arrhythmias, and thromboembolic cerebrovascular events (Jefferies 2013).

Treatment: There is no cure for Barth syndrome. Supportive therapy includes physiotherapy, antibiotics for any bacterial infections, heart function monitoring, and medications if necessary. In severe cases, heart transplant might be required (Spencer et al. 2006).

Cohen syndrome

Description: Cohen syndrome (OMIM# 216550) is an autosomal recessive disorder characterized by developmental delay, microcephaly, typical facial features, ophthalmologic abnormalities, and granulocytopenia. Facial features include high-arched or wave-like eyelids, short philtrum, and thick hair with a low hairline. Cohen syndrome patients often develop retinchoroidal dystrophy and myopia by 5 years of age. Other manifestations include joint laxity, narrow hands and feet, sandal-gap toes, and a sociable disposition (Kivitie-Kallio and Norio 2001).

Pathogenesis: Cohen syndrome is caused by mutations in the *COH1* gene on chromosome 8 that might be involved in vesicle-mediated sorting and intracellular protein transport (Kolehmainen et al. 2003).

Immune abnormalities: The neutropenia in Cohen syndrome, thought to be secondary to a defect in myeloid differentiation, is typically intermittent and usually does not predispose patients to infections (Kivitie-Kallio and Norio 2001).

CVS abnormalities: Some patients suffer from mitral valve prolapse and decreasing left ventricle function with age (Sack and Friedman 1980; Mehes et al. 1988; Kivitie-Kallio et al. 1999).

Treatment: The retinopathy requires regular monitoring and eyeglasses. The neutropenia does not require any specific treatment (Kivitie-Kallio and Norio 2001).

Shwachman–Diamond syndrome

Description: Shwachman–Diamond syndrome (SDS) is an autosomal recessive disorder (OMIM# 260400) characterized by neutropenia, severe exocrine pancreatic insufficiency, liver abnormalities, and bone marrow failure or myelodysplastic syndrome. Additionally, patients are predisposed to develop leukemia (Dror et al. 2011). SDS is often associated with neuro-developmental abnormalities and metaphyseal dysplasia.

Pathogenesis: SDS is caused by mutations in the *SBDS* gene on chromosome 7. Although the precise function of the protein encoded by the *SBDS* gene is unknown, it is thought to be involved in ribosome synthesis (Sen et al. 2011).

Immune abnormalities: SDS patients are prone to recurrent bacterial and viral infections, especially otitis media, pneumonia, septicemia, osteomyelitis, and skin infections. The neutropenia usually presents at an early age and can be either intermittent or chronic. Some patients with SDS have variable immune abnormalities (Dror et al. 2001). These include low IgG or IgG subclasses, low percentage of circulating B cells, decreased in vitro B-cell proliferation, and a lack of specific antibody production. T lineage abnormalities among these patients comprise a low percentage of total circulating T cells or CD3⁺/CD4⁺ T-cell subpopulations and decreased in vitro T-cell proliferation. Additionally, chemotaxis of the neutrophils might be reduced.

CVS abnormalities: Ventricular and atrial septal defects, patent foramen ovale, and patent ductus arteriosus may occur in 19% of SDS patients (Myers et al. 2014). Heart failure, possibly secondary to dilated cardiomyopathy, is another common cardiac manifestation in SDS, and it can be severe enough to cause death in the neonatal period (Toiviainen-Salo et al. 2008; Kopel et al. 2011). Histopathology in those cases show myocardial necrosis or fibrosis (Toiviainen-Salo et al. 2008; Burroughs et al. 2009).

Treatment: SDS patients require close monitoring throughout their lifetime for malignancy. Oral pancreatic enzyme supplementation may be necessary, especially early in life. HSCT should be considered in SDS patients with bone marrow failure, myelodysplastic syndrome, and leukemia. Cardiotoxicity, including transient congestive heart failure and long-term cardiac hypokinesia, can occur also following HSCT (Burroughs et al. 2009).

WHIM syndrome

Description: Warts, hypogammaglobulinemia, infections, and myelokathexis (WHIM) syndrome is an autosomal dominant disorder characterized by difficult-to-treat warts, low IgG levels, recurrent bacterial infections, and neutropenia (Kawai and Malech 2009).

Pathogenesis: WHIM syndrome is caused by mutations in the *CXCR4* chemokine receptor gene (OMIM# 193670). This gene is part of a pathway involved in regulation of hematopoiesis, organogenesis, bone marrow homeostasis, and leukocyte motion (Badolato et al. 2012). Mutations in the terminal domain of *CXCR4* enhance retention of mature hyper-segmented neutrophils in the bone marrow leading to neutropenia (Kawai and Malech 2009).

Immune abnormalities: The key immunologic features of WHIM syndrome include severe neutropenia. During infections, stress, or granulocyte colony stimulating factor administration, neutrophil numbers in the peripheral blood temporarily increase. WHIM patients might have reduced T- and B-cell numbers as well as hypogammaglobulinemia and abnormal antibody production (Kawai and Malech 2009). Patients with WHIM syndrome are particularly susceptible to papilloma virus infections, leading to warts that can be very extensive and progress to carcinoma (Kawai and Malech 2009).

CVS abnormalities: Few patients with WHIM syndrome have been reported with structural cardiac defects, such as tetralogy of Fallot, possibly because of a role of *CXCR4* in cardiac embryogenesis (Badolato et al. 2012).

Treatment: Antibiotic prophylaxis and immunoglobulin replacement can help prevent infections. Papilloma lesions should be aggressively treated and carefully monitored for malignant changes (Kawai and Malech 2009).

MonoMAC syndrome

Description: Several groups recently identified monocytopenia with predisposition to nontuberculosis mycobacterial infections among patients with autosomal dominant mutations in the zinc finger transcription factor *GATA2* (OMIM# 137295) (Hsu et al. 2011; Dickinson et al. 2011). Patients are also prone to develop

myelodysplastic syndrome and acute myeloid leukemia (Ostergaard et al. 2011).

Pathogenesis: *GATA2* is expressed in hematopoietic progenitors, including early erythroid cells, mast cells, megakaryocytes, and also in nonhematopoietic embryonic stem cells, possibly explaining the common hematological abnormalities observed among patients (Hsu et al. 2011).

Immune abnormalities: Patients typically have markedly reduced number of monocytes, B cells, and NK cells in the peripheral blood, resulting in susceptibility to mycobacteria as well as opportunistic fungal infections and disseminated human papillomavirus infections. Abnormal macrophage function in tissue might contribute to the frequent development of pulmonary alveolar proteinosis in these patients. IgG levels and ex vivo T-cell functions are often normal. Erythema nodosum and autoimmunity can be the presenting features (Hsu et al. 2011; Dickinson et al. 2011).

CVS abnormalities: Patients also suffer from primary lymphedema, possibly reflecting the important role of *GATA2* in controlling capillary network formation (Hsu et al. 2011, Johnson et al. 2012; Kazenwadel et al. 2012).

Treatment: Patients should be monitored closely for the development of opportunistic infections as well as myelofibrosis and leukemia. HSCT has been curative in some cases.

Chronic granulomatous disease

Description: Chronic granulomatous disease (CGD) is characterized by inefficient intracellular killing of pathogens resulting in recurrent, severe life threatening infections, inflammation, and granuloma formation (Jones et al. 2008).

Pathogenesis: CGD is caused by mutations in one of 5 genes *CYBB* (OMIM# 306400), *CYBA* (OMIM# 608508), *NCF1* (OMIM# 608512), *NCF2* (OMIM# 608515), and *NCF4* (OMIM# 601488) resulting in absent or nonfunctional proteins, gp91 phox, p22 phox, p47 phox, p67 phox, and p40 phox, respectively. Mutations in the *CYBB* gene cause X-linked CGD and occur in about two-thirds of the patients. All other gene mutations are associated with autosomal recessive CGD. These mutations impair the function of the nicotinamide adenine dinucleotide phosphate-oxidase enzyme complex and prevent production of reactive

oxidative species. The impaired oxidative response also enables rapid identification of CGD by flow cytometry analysis, as neutrophils display reduced or absent oxidative burst following ex vivo stimulation (Jones et al. 2008).

Immune abnormalities: Patients are particularly prone to infections by catalase-positive pathogens such as *Staphylococcus aureus*, *Burkholderia cepacia*, *Serratia marcescens*, *Nocardia*, *Aspergillus*, *Salmonella*, and *Bacillus Calmette-Guerin*. Patients also suffer from persistent inflammation and autoimmunity, particularly involving the gastrointestinal tract, as well as formation of multiple granulomas, which may even cause obstruction of viscera such as the stomach, urinary track, etc. (Marciano et al. 2004; Jones et al. 2008).

CVS abnormalities: A large retrospective chart review of CVS disease among 229 patients with CGD followed at the U.S. National Institute of Health, identified 2 patients with coronary artery disease and another patient with aortic and mitral valve regurgitations (Leiding et al. 2013). Interestingly, the 3 patients with CVS abnormalities had p47^{PHOX} mutations. One of these patients was also found to have cerebral aneurysms, diagnosed during family screening. Affected patients had other risk factors for CVS abnormalities such as diabetes and dysregulated cholesterol; hence, the direct contribution of defective reactive oxidative species production in CVS disease development needs to be further studied. Interestingly, a recent study identified increased nitric oxide, a potent vasodilator molecule produced by endothelial cells, and significantly higher flow-mediated dilatation among 30 patients with CGD (Loffredo et al. 2013). The clinical significance of these findings for the development of CVS abnormalities is still not clear.

Treatment: Patients with CGD should receive antibacterial and antifungal prophylaxis. Infections and granulomas should be treated promptly with antibiotics and immunosuppressive medications, respectively. HSCT, from family related or unrelated HLA-matched donors, can cure CGD. Currently, human trials of gene therapy for CGD are underway.

Well-defined syndromes with immunodeficiency

DiGeorge syndrome

Description: DGS, estimated to occur in 1:3000 live births, is characterized by variable combinations of

neonatal hypocalcaemia, thymus hypoplasia, and conotruncal defects that disrupt the cardiac outflow tract (OMIM# 188400). De novo or autosomal dominant inheritance deletion of chromosome 22q11.2 are found among 80%–90% of the patients, although haploinsufficiency of others chromosomes such as 10p13 or 4q21.3-q25 have also been implicated. Other features commonly identified among patients with 22q11.2 microdeletion include renal anomalies, ophthalmologic abnormalities, developmental delay, learning disabilities, schizophrenia, and bipolar disorders as well skeletal defects and short stature. Palate abnormalities are also common, resulting in velocardiofacial syndrome (OMIM#192430). Patients also have typical dysmorphic facial features including micrognathia, low set ears, telecanthus with short palpebral fissures, upward or downward slanting eyes with short philtrum, and a relatively small mouth (Kobrynski and Sullivan 2007).

Pathogenesis: DGS is thought to be secondary to disrupted cervical neural crest migration into the derivatives of the 3rd and 4th pharyngeal arches and pouches. Among patients with 22q11.2 deletion, haplo-insufficiency of the *TBX1* gene, which encodes a transcription factor important for neural crest cells migration, is thought to cause the majority of the features (Kobrynski and Sullivan 2007).

Immune abnormalities: DGS is the most common human T-cell immune deficiency. The abnormalities of the thymus impair thymocytes development and selection, often manifesting with diminished T-cell counts in the peripheral blood, although the responses of T cells to stimulation is often preserved. Indeed in the majority of patients with DGS, the laboratory T-cell abnormalities are not associated with increased susceptibility to infections. Moreover, the number of T cells increases during infancy and childhood among most patients. “Complete DGS” with thymus aplasia and practically absent T cells has been reported in <1% of the patients. However, because of the potential severity and lethality of this condition, as well as the lack of correlation with nonimmune abnormalities, complete DGS should be excluded in all patients. Severe thymus defects cause marked reduction of new T-cell production, resulting in low TREC. Importantly, TREC, which can be measured in peripheral blood using PCR, enable detection of severe DGS in newborn screening programs recently implemented among many counties. Severe DGS can also manifest as Omenn syndrome with misleading normal or even elevated T-cell numbers. Few patients have also been identified with hemophagocytic

lymphohistiocytosis. Autoimmune conditions, resulting in hematological cytopenias or rheumatologic complications, are also common in patients with DGS. Infrequently, patients with DGS also display humoral immunodeficiency with reduced production of IgG or specific antibodies (Kobrynski and Sullivan 2007; Gennery 2012).

CVS abnormalities: Approximately 80% of patients with DGS and 22q11.2 deletion syndrome have various congenital heart defects (Momma 2010). The heart disease typically manifests as cardiac outflow tract anomalies related to impaired embryogenesis. The most common defects are tetralogy of Fallot, ventricular septal defect, interrupted aortic arch, and truncus arteriosus. Other reported anomalies include right aortic arch and aberrant right subclavian artery, atrial septum defect, pulmonary valve stenosis, double outlet right ventricles, coarctation of the aorta, great arteries transposition, etc. (Momma 2010).

Treatment: Management of DGS depends on the clinical phenotype. Cardiac defects may require early surgical intervention, whereas the hypocalcaemia often can be treated with calcium supplementation (Kobrynski and Sullivan 2007). Antimicrobial prophylaxis and immunoglobulin replacement should be considered for the immunodeficiency (Gennery 2012) as well as adherence to cytomegalovirus-negative irradiated blood products. Yet for most patients with DGS, the normal or near-normal immune system does enable administration of live viral vaccines (Al-Sukaiti et al. 2010). Allogeneic thymus tissue transplantation or HSCT have been used for the rare “complete DGS” with variable success.

CHARGE syndrome

Description: A multiple congenital cardiac and immune anomaly disorder (OMIM# 214800) characterized by coloboma of the eye, heart anomalies, atresia choanae, retardation of mental and physical development, genital anomalies, and ear abnormalities/deafness is known as CHARGE. There is much phenotypic variability among patients with CHARGE syndrome, and the clinical diagnostic criteria have been revised several times (Bergman et al. 2011).

Pathogenesis: CHARGE syndrome typically arises from autosomal dominant mutations in transcription regulator *chromodomain helicase DNA-binding protein-7* gene. During embryonic development, the

product of this gene is thought to be essential for normal formation of multipotent migratory neural crest cells. Further, its expression in the central nervous system, semicircular canals, and neural crest of the pharyngeal arches explains some of the diverse tissue abnormalities found in CHARGE syndrome (Gennery et al. 2008; Bergman et al. 2011).

Immune abnormalities: Mild to moderate T-cell deficiencies, including Omenn syndrome, are infrequently observed among patients with CHARGE syndrome. Isolated antibody deficiency is also rarely found in CHARGE syndrome (Writzl et al. 2007; Gennery et al. 2008; Jyonouchi et al. 2009).

CVS abnormalities: Congenital heart lesions occur in 75%–80% of patients (Blake and Prasad 2006). Conotruncal heart defects, especially tetralogy of Fallot, are the most common manifestation, similar to DGS. However, many lesions have been reported including atrioventricular canal defects, aortic arch defects, and patent ductus arteriosus, as well as atrial and ventricular septal defects (Blake and Prasad 2006).

Treatment: Surgical intervention may be required for choanal atresia and cardiac defects. All CHARGE syndrome patients should have ophthalmological follow-up as well as hearing screens and hearing aids if necessary. Potential immunodeficiency should be screened for and managed accordingly (Blake and Prasad, 2006).

Wiskott–Aldrich syndrome

Description: Wiskott–Aldrich syndrome (WAS) is an X-linked disorder characterized by thrombocytopenia, severe eczema, and recurrent infections (OMIM# 301000). Thrombocytopenia may manifest early as petechiae, bruising, bloody diarrhea, or excessive bleeding after circumcision. The platelets are typically small in size (Ochs et al. 2009).

Pathogenesis: WAS is caused by mutations in the WAS gene located on the X chromosome. Accordingly, it is predominantly found in males, although rarely females with nonrandom X inactivation may develop a similar phenotype. The WAS gene codes for the WAS protein, a cytoskeletal regulator expressed in hematopoietic cells (Ochs et al. 2009; Bouma et al. 2009). WAS protein is involved in cell signaling, migration, and immune synapse formation (Ochs et al. 2009; Bouma et al. 2009). Many different types of mutations within this gene cause WAS; however, clear correlation

between the mutations and phenotypic severity have not been established.

Immune abnormalities: Infections, including recurrent otitis media, bacterial pneumonias, and skin infections are common among patients suffering from WAS (Ochs et al. 2009). Opportunistic infections such as *Pneumocystis jiroveci* pneumonia can also occur, reflecting the combined T- and B-cell dysfunction in this condition. There is often decreased numbers of circulating T cells, particularly CD8⁺ T cells, as well as diminished T-cell function. IgG and IgM may be normal or decreased, whereas IgA and IgE are typically elevated (Ochs et al. 2009). Infrequently, elevated IgM and reduced IgG, reminiscent of hyper-IgM syndrome can be found as well as abnormal responses to polysaccharide antigens (Ochs et al. 2009). Impaired immune cell migration and defects in the innate immune system are also observed in WAS (Bouma et al. 2009). Other manifestations of this syndrome include autoimmune disorders, such as hemolytic anemia or immune-mediated thrombocytopenia, which further complicates the diagnosis and management of the platelet disorder (Ochs et al. 2009). Increased susceptibility to malignancies, including lymphoma and leukemia, as well as solid tumors is noted.

CVS abnormalities: Aneurysm formation as well as small, medium, and large (including aorta) vessel vasculitis are found in few patients with WAS. These include a 24-year-old patient who developed necrotizing vasculitis of small and medium sized arteries, resulting in aneurysms of his renal, hepatic, and superior mesenteric arteries (McCluggage et al. 1999) and a 7-year-old boy with aortic root dilatation (Ono et al. 2009). Pathology in the latter child showed aortitis with inflammatory infiltration and necrotic areas. Additional WAS patients with aortic aneurysms have been described (Pellier et al. 2011). The pathogenesis for the vasculitis is not clear, yet it might precede the thrombocytopenia (Mahlaoui et al. 2013).

Treatment: Patients with WAS typically require antimicrobial prophylaxis to prevent infections and platelet transfusions to treat significant bleeding episodes (Ochs et al. 2009). Immunosuppressive therapy may be necessary to treat autoimmune manifestations particularly the thrombocytopenia (Ochs et al. 2009). Splenectomy was previously used for severe thrombocytopenia; however, such procedure increases the risk for infections and it does not prevent malignancy. HSCT, from related or unrelated HLA-matched donors has been

shown to be highly beneficial for patients suffering from WAS, particularly if performed at an early age. Currently, human trials of gene therapy for WAS are underway.

Roifman syndrome

Description: Roifman syndrome (OMIM# 300258) is characterized by antibody deficiency, spondyloepiphyseal dysplasia, growth retardation, and retinal dystrophy (Roifman 1999). Associated manifestations include intellectual disability, dysmorphic features, and hypogonadism.

Pathogenesis: The gene responsible for Roifman Syndrome has not yet been identified.

Immune abnormalities: Roifman Syndrome is associated with humoral immunodeficiency, particularly poor specific antibody response. This results in recurrent sinopulmonary infections including otitis media and pneumonia (Roifman 1999).

CVS abnormalities: A 14-year-old boy with Roifman Syndrome was reported to suffer from left ventricular noncompaction and heart failure (Mandel et al. 2001).

Treatment: Patients are managed with immunoglobulin replacement and close monitoring.

Schimke immuno-osseus dysplasia

Description: Schimke immuno-osseus dysplasia (SIOD) is an autosomal recessive disorder characterized by spondyloepiphyseal dysplasia, immunodeficiency, facial dysmorphism, and progressive renal disease (OMIM# 242900). Other features include skin lesions, hypothyroidism, headaches, cerebral ischemia, and enteropathy (Boerkoel et al. 2000). There appears to be 2 forms of SIOD: juvenile onset and the more severe infantile onset (Boerkoel et al. 2000).

Pathogenesis: SIOD is caused by biallelic loss-of-function mutations in the *SMARCA1* gene (Clewing et al. 2007). This gene encodes a protein involved in chromatin remodeling, but the pathogenic mechanism of SIOD is still unclear (Clewing et al. 2007).

Immune abnormalities: Immune dysfunction in SIOD is primarily T-cell deficiency and dysfunction resulting in increased susceptibility to opportunistic

fungal, viral, and bacterial infections (Boerkoel et al. 2000; Lev et al. 2009).

CVS abnormalities: Vascular abnormalities occur in many patients, including transient ischemic attacks and (or) cerebral ischemia, putatively secondary to early onset, and progressive arteriosclerosis (Boerkoel et al. 2000; Clewing et al. 2007). A recent study among more than 50% of SIOD patients found arteriosclerosis associated with vascular intimal and medial hyperplasia, smooth muscle cell hyperplasia, and fragmented and disorganized elastin fibers (Morimoto et al. 2012). Few patients were reported to have cerebral artery stenosis and *moya-moya* phenomenon (Boerkoel et al. 1998).

Treatment: Dialysis and renal transplant are used for the progressive renal failure (Boerkoel et al. 2000).

Hyper-IgE syndromes

Description: Hyper-IgE Syndrome (HIES) represents a group of disorders characterized by recurrent skin and lung infections, eczema, and elevated IgE levels (Heimall et al. 2010). An autosomal dominant form (OMIM# 147060), also known as Job's Syndrome, is associated with distinctive coarse facial features, delayed shedding of primary teeth, frequent bone fractures, spinal deformities, vascular anomalies, and development of lymphoma (Heimall et al. 2010). Pulmonary infections often result in pneumatoceles and (or) bronchiectasis. The autosomal recessive form (OMIM# 243700) is often associated with a more profound immunodeficiency resulting in susceptibility to viral infections, particularly papilloma and herpes viruses, autoimmunity, sclerosing cholangitis, and vasculitis (Heimall et al. 2010; Alsum et al. 2013).

Pathogenesis: Autosomal dominant HIES can be caused by mutations in the *STAT3* gene (Heimall et al. 2010). The *STAT3* protein is part of the JAK-STAT signal-transducing pathway, responsible for activating gene transcription and translation (Heimall et al. 2010). Autosomal recessive HIES is caused primarily by mutations in *DOCK8*, although in a few patients defects in *TYK2* were identified. Somatic mutations in *STAT3* have also been recently identified among patients with a milder phenotype (Hsu et al. 2013).

Immune abnormalities: Markedly elevated IgE levels are common among patients with HIES; however, similar levels might be found among patients with atopic conditions as well as other immune deficiencies.

Patients with HIES might also have fluctuating as well as normal or near normal IgE levels, making the IgE level an unreliable diagnostic criteria. Some patients might have impaired antibody responses or decreased IgG levels. The autoimmunity in HIES is attributed to decreased IL-17 producing T-helper cells. Importantly, severe T- and B-cell defects can occur early in life in some patients with *DOCK8* mutations.

CVS abnormalities: Vascular abnormalities involving arteries or veins of the brain, lungs, heart, and skin as well as coronary and aortic aneurysms have been identified among patients with *STAT3* defects (Yavuz and Chee, 2010). Similar vascular abnormalities including giant aortic aneurysm, brain infarction, and *moya-moya* syndrome were identified in patients with *DOCK8* mutations (Alsum et al. 2013). It is still not clear whether the vascular lesions are secondary to congenital angiogenesis defects post-infectious inflammation or to autoimmunity.

Treatment: Patients with abnormal antibody production should receive immunoglobulin replacement. Patients with severe autosomal recessive HIES can benefit from HSCT.

Immunodeficiency with centromeric instability and facial anomalies

Description: Immunodeficiency with centromeric instability and facial anomalies (ICF) syndrome (OMIM# 242860) is an autosomal recessive disease. ICF can be caused by mutations in the *DNA methyltransferase 3B* gene at chromosome 20q11.2 (designated ICF1) or the *zinc-finger and BTB domain-containing 24* gene (ICF2). Patients have mild facial anomalies including a round face, flat nasal bridge, hypertelorism, epicanthus, up-turned nose, macroglossia, micrognathia, and low-set ears. The majority of ICF patients have delayed walking and speech, and some may suffer from intellectual disabilities. Diagnosis of ICF can be established by demonstrating excess deletions and peri-centromeric breaks of chromosomes 1, 9, and 16 in metaphases of stimulated peripheral blood lymphocytes (Weemaes et al. 2013).

Pathogenesis: Hypomethylation of various genes, including those important for immune and neurological development, as well as satellite DNA located in the pericentromeric regions of chromosomes 1, 9, and 16

may lead to the diverse clinical and laboratory features (Weemaes et al. 2013).

Immune abnormalities: Severe infections including pneumonia, sepsis, as well as opportunistic infections caused by pathogens such as *Candida albicans*, *Pneumocystis jiroveci*, and JC virus may occur in ICF patients. Hypo- or agammaglobulinemia is found among most patients with ICF. Patients' circulating B cells contain an increased proportion of immature cells and reduced memory cells. Additionally, B cells are more prone to undergo apoptosis upon in vitro activation. The numbers of CD3⁺, CD4⁺, and CD8⁺ cells are usually normal. However, with age, some patients exhibit T-cell lymphopenia and reduced proliferation of T cells following ex vivo stimulation. Few patients also develop neutropenia in the second decade of life (Weemaes et al. 2013).

CVS abnormalities: Ventricular and atrial septum defects were reported in few patients.

Treatment: Antibody deficiency should be treated with immunoglobulin replacement. HSCT was performed in few patients because of severe infections or myelodysplasia with some success (Weemaes et al. 2013).

Hepatic veno-occlusive disease with immunodeficiency

Description: Hepatic veno-occlusive disease with immunodeficiency (VODI) syndrome (OMIM# 235550) was first described in Lebanese descendants in Australia, followed by descriptions from other continents and populations. VODI is an autosomal recessive disease caused by mutations in the *SP110* gene (Roscioli et al. 2006). Some patients suffer from microcephaly, leukodystrophy, multiple small cerebral calcifications, and neuro-developmental defects, as well as pulmonary fibrosis, hepatosplenomegaly, and thrombocytopenia. Histological and ultrasound-Doppler can establish the diagnosis of hepatic sinusoidal obstructive syndrome.

Pathogenesis: The molecular pathogenesis of VODI is not clear; however, it might be related to abnormalities in the PML nuclear body protein, a transcriptional factor that enhances transcription of genes with retinoic acid response elements (Roscioli et al. 2006).

Immune abnormalities: Multiple infections with pathogens such as *Pneumocystis jiroveci*, cytomegalovirus, and *Candida* often occur in infancy, reflecting a combined T- and B-cell immunodeficiency. Severe hypogammaglobulinemia as well as reduced IgM and IgA are also commonly found. Histological evaluations often reveal absent lymph nodes' germinal centers and tissue plasma cells. Neutropenia can also occur (Roscioli et al. 2006).

CVS abnormalities: In addition to the characteristic hepatic veno-occlusive disease, left atrial endocardial fibrosis can occur (Roscioli et al. 2006).

Treatment: Patients should receive immunoglobulin replacement and *Pneumocystis jiroveci* prophylaxis. HSCT was successful among few patients (Ganaiem et al. 2013).

Facial dysmorphism, immunodeficiency, livedo, and short stature (FILS syndrome)

Description: Recently, a single large consanguineous French kindred was identified with mild facial dysmorphism, immunodeficiency, livedo, and short stature caused by mutations in the DNA polymerase ϵ gene (*POLE*) (Pachlopnik Schmid et al. 2012). Few patients also display bone dysplasia with lacunar lesions, cortical thickening, and modeling defects of long bone diaphysis as well as metaphyseal striae. Growth impairment can be observed during early childhood impairing adult height.

Pathogenesis: The precise mechanism is still not clear; however, it is hypothesized that absence of DNA polymerase ϵ impedes proliferation and G1- to S-phase progression in lymphocytes, chondrocytes, and osteoblasts (Pachlopnik Schmid et al. 2012).

Immune abnormalities: The immunodeficiency often manifests early in life with recurrent respiratory tract infections resulting in bronchiectasis. Some patients also suffer recurrent meningitis caused by *Streptococcus pneumoniae*. Immune evaluation of the patients shows decreased IgM and IgG2 levels, reduced isohemagglutinin titers, and a predominant lack of antibodies to polysaccharide antigens. Patients may have low memory B- and naive T-cell counts as well as decreased T-cell proliferation (Pachlopnik Schmid et al. 2012).

CVS abnormalities: Livedo on the cheeks, forearms, and (or) legs is present in most patients and can sometimes be noticed from birth. With increasing age, telangiectasia can be observed on the cheeks (Pachlopnik Schmid et al. 2012).

Treatment: Patients might benefit from prophylactic antibiotics. Whether immunoglobulin replacement is beneficial for these patients is not clear.

Disease of immune dysregulation

X-linked lymphoproliferative disease

Description: X-linked lymphoproliferative disease (XLP) is characterized by severe immune dysregulation, often triggered by EBV infection (OMIM# 308240). Some patients suffer recurrent infections, whereas others might present with fulminant infectious mononucleosis and liver failure as well as EBV or non-EBV induced hemophagocytic lymphohistiocytosis and lymphoma (Booth et al. 2011). The phenotype is highly variable, even within the same family. Other less common presenting features include aplastic anemia, vasculitis, colitis, and chronic gastritis (Booth et al. 2011).

Pathogenesis: XLP disease is caused by mutations in the *SH2D1A* gene, located on the X chromosome; hence, the disease affects males. The gene encodes SAP, a protein that helps regulate immune function in T, NK, and possibly B cells (Booth et al. 2011).

Immune abnormalities: In XLP, the numbers of circulating T cells are normal, but their function is impaired. Some patients display hypogammaglobulinemia or dysgammaglobulinemia. Low numbers of NK cells are also common (Booth et al. 2011).

CVS abnormalities: Systemic vasculitis has been reported in few patients with XLP. These include small and medium vessel multi-organ lymphocytic vasculitis (Dutz et al. 2001).

Treatment: Antibody deficiency should be treated with immunoglobulin replacement. Because of the risk for hemophagocytic lymphohistiocytosis and lymphoma, preventive HSCT with appropriate donors should be considered, particularly in young patients (Booth et al. 2011).

FADD deficiency

Description: FADD deficiency (OMIM# 613759) can cause autoimmune lymphoproliferative syndrome (ALPS) (Bolze et al. 2010). Patients suffer recurrent bacterial and viral infections, hepatic dysfunction, functional asplenia, encephalopathy, and cardiac malformations.

Pathogenesis: FADD deficiency is an autosomal recessive disease caused by mutations in the *FADD* gene on chromosome 11q13.3 (Bolze et al. 2010). The *FADD* gene encodes an adaptor molecule that interacts with FAS, an apoptosis-inducing receptor molecule. The FADD protein is also involved in autophagy, inflammation, innate immunity, cell proliferation, and tumor development. Mutations in *FADD* lead to impaired FAS-dependent apoptosis and immune dysregulation (Bolze et al. 2010).

Immune abnormalities: FADD deficiency is associated with elevated double negative T cells, and elevated IL-10 and Fas-Ligand levels. FADD-deficient patients display Howell–Jolly bodies in their blood smears, indicating functional asplenia, which may explain the susceptibility of patients to severe and invasive bacterial infections (Bolze et al. 2010).

CVS abnormalities: Two of the 4 patients with FADD deficiency had cardiac malformations. One had pulmonary atresia with a ventricular septal defect and the other had a left-sided superior vena cava, draining into the left atrium (Bolze et al. 2010). Mice are known to express FADD in the myocardium and mice with FADD deficiency have embryonic defects, suggesting that FADD deficiency might affect cardiogenesis (Bolze et al. 2010).

Treatment: At present there is no curative therapy for this syndrome. Only one of the 4 described patients is still alive (Bolze et al. 2010).

Aicardi–Goutières syndrome

Description: Aicardi–Goutières syndrome (AGS) is characterized by familial encephalopathy resembling intrauterine viral infection (OMIM# 225750). Patients suffer from varying severities of psychomotor retardation, microcephaly, dystonic posturing, and spasticity. Chilblain-like skin lesions and congenital glaucoma may also be present. Brain imaging often reveals intracerebral calcifications, particularly in the basal ganglia,

cerebral atrophy, and leukodystrophy. The syndrome manifests in infancy, and death often occurs early in childhood; however, milder forms and survival into adulthood can occur. AGS is caused by mutations in different genes including *TREX1*, *RNASEH2A*, *RNASEH2B* and *RNASEH2C*, as well as *ADAR1* and *SAMHD1* (Kavanagh et al. 2008).

Pathogenesis: AGS is thought to be caused by impaired clearance of nucleic acids and abnormal intracellular accumulation of single-stranded DNA species, which trigger an inappropriate type I interferon-mediated immune response leading to vasculitis and vascular changes (Kavanagh et al. 2008).

Immune abnormalities: Patients may suffer from various autoimmune manifestations including arthritis, thrombocytopenia, and elevated transaminases, and they may also develop autoantibodies. Typically lymphocytosis and elevated interferon- α can be found in the cerebrospinal fluid, mimicking congenital transplant-acquired infection, which needs to be excluded. Pterins in cerebrospinal fluid can help establish the diagnosis (Kavanagh et al. 2008).

CVS abnormalities: Defects in *TREX1* are associated with transient ischemic attacks and strokes (Kavanagh et al. 2008). Histological assessments show obliterative vasculopathy in the brain, whereas fluorescein angiograms demonstrate retinal vasculopathy. Additionally, small vessel gastrointestinal bleeding and renal arterio-
lonephrosclerosis have been observed. Few patients with *SAMHD1* deficiency have stenosis of carotid and intracranial vessels leading to stroke (Kavanagh et al. 2008).

Treatment: Recognizing AGS as an autoimmune/autoinflammatory condition has led to the use of various immune modulating agents, although it is difficult to assess their efficacy. Alternative strategies exploring blocking antibodies to interferon- α , reverse transcriptase inhibition, or B-cell depletion are currently being developed (Crow et al. 2014).

Autoinflammatory disorders

HOIL1 deficiency

Description: Mutations in *HOIL1* (*RBCK1*) can cause an autosomal recessive disease characterized by persistent infections, auto-inflammation, severe failure to

thrive, liver fibrosis with hepatocytes vacuoles, and amylopectin-like deposits (Boisson et al. 2012).

Pathogenesis: Three patients from 2 unrelated families were shown to carry loss-of-function mutations in *HOIL1*, a component of the linear ubiquitination chain assembly complex. The defect impairs fibroblasts NF- κ B activation in response to IL-1 β and TNF- α . In contrast, the patients' mononuclear leukocytes, particularly monocytes, were hyper-responsive to IL-1 β .

Immune abnormalities: Patients are susceptible to pyogenic bacterial and CMV infections. Auto-inflammation may manifest in infancy with recurrent fevers, severe eczematous rash, lymphadenopathy, hepato-splenomegaly, inflammatory bowel disease, and elevated inflammatory cytokines. Laboratory evaluations demonstrate increased IgA, decreased memory B cells and minor defects in antibody production following vaccinations (Boisson et al. 2012).

CVS abnormalities: Patients display muscular amylopectinosis, evident already in infancy, which contributes to progressive muscular atrophy as well as severe dilated cardiomyopathy and congestive cardiac failure. Amylopectin-like deposits are found in cardiomyocytes as well as gut muscular mucosa and skeletal muscle cells (Boisson et al. 2012).

Treatment: Supportive therapy for infections and immune dysregulation is necessary. One patient, who died 3 years after successful HSCT, had sub-endocardial fibrosis with cytoplasmic vacuolization of myocardiocytes and modest mononuclear cell infiltrates, indicative of chronic myocarditis (Boisson et al. 2012).

Conclusion

In conclusion, as described in this comprehensive review, the expanding spectrum of PID requires increased alertness to the possibility of CVS involvement as an important contributor to the diagnosis and management of these patients.

REFERENCES

- Abdollahpour, H., Appaswamy, G., Kotlarz, D., Diestelhorst, J., Beier, R., Schaffer, A.A., Gertz, E.M., Schambach, A., Kreipe, H.H., Pfeifer, D., Engelhardt, K.R., Rezaei, N., Grimbacher, B., Lohrmann, S., Sherkat, R., and Klein, C. 2012. The phenotype of human STK4 deficiency. *Blood*. **119**(15):3450–3457. PMID: 22294732. doi: 10.1182/blood-2011-09-378158.
- Albert, M.H., Gennery, A.R., Greil, J., Cale, C.M., Kalwak, K., Kondratenko, I., Mlynarski, W., Notheis, G., Führer, M., Schmid, I., and Belohradsky, B.H. 2010. Successful SCT for Nijmegen breakage syndrome. *Bone Marrow Transpl.* **45**(4):622–626. PMID: 19684627. doi: 10.1038/bmt.2009.207.
- Aleman, K., Noordzij, J.G., De Groot, R., Van Dongen, J.J., and Hartwig, N.G. 2001. Reviewing Omenn syndrome. *Eur. J. Pediatr.* **160**(12):718–725. PMID: 11795679. doi: 10.1007/s004310100816.
- Al-Sukaiti, N., Reid, B., Lavi, S., Al-Zaharani, D., Atkinson, A., Roifman, C.M., and Grunebaum, E. 2010. Safety and efficacy of measles, mumps, and rubella vaccine in patients with DiGeorge syndrome. *J. Allergy Clin. Immunol.* **126**(4):868–869. PMID: 20810153. doi: 10.1016/j.jaci.2010.07.018.
- Alsum, Z., Hawwari, A., Alsmadi, O., Al-Hissi, S., Borrero, E., Abu-Staiteh, A., Khalak, H.G., Wakil, S., Eldali, A.M., Arnaout, R., Al-Ghonaum, A., Al-Muhsen, S., Al-Dhekri, H., Al-Saud, B., and Al-Mousa, H. 2013. Clinical, immunological and molecular characterization of DOCK8 and DOCK8-like deficient patients: single center experience of twenty-five patients. *J. Clin. Immunol.* **33**(1):55–67. PMID: 22968740. doi: 10.1007/s10875-012-9769-x.
- Badolato, R., Dotta, L., Tassone, L., Amendola, G., Porta, F., Locatelli, F., Notarangelo, L.D., Bertrand, Y., Bachelier, F., and Donadieu, J. 2012. Tetralogy of fallot is an uncommon manifestation of warts, hypogammaglobulinemia, infections, and myelokathexis syndrome. *J. Pediatr.* **161**(4):763–765. PMID: 22748845. doi: 10.1016/j.jpeds.2012.05.058.
- Banka, S., Chervinsky, E., Newman, W.G., Crow, Y.J., Yeganeh, S., Yacobovich, J., Donnai, D., and Shalev, S. 2011. Further delineation of the phenotype of severe congenital neutropenia type 4 due to mutations in G6PC3. *Eur. J. Hum. Genet.* **19**(1):18–22. PMID: 20717171. doi: 10.1038/ejhg.2010.136.
- Bastianon, V., Giglioni, E., Businco, L., Fiorilli, M., and Chessa, L. 1993. Cardiac anomalies in ataxia-telangiectasia. *Am. J. Dis. Child.* **147**(1):20–21. PMID: 8418594. doi: 10.1001/archpedi.1993.02160250022008.
- Begin, P., Patey, N., Mueller, P., Rasquin, A., Sirard, A., Klein, C., Haddad, É., Drouin, É., and Le Deist, F. 2013. Inflammatory bowel disease and T cell lymphopenia in G6PC3 deficiency. *J. Clin. Immunol.* **33**(3):520–525. PMID: 23180359. doi: 10.1007/s10875-012-9833-6.

- Bergman, J.E.H., Janssen, N., Hoefsloot, L.H., Jongmans, M.C.J., Hofstra, R.M.W., and Van Ravenswaaij-Arts, C.M.A. 2011. CHD7 mutations and CHARGE syndrome: the clinical implications of an expanding phenotype. *J. Med. Genet.* **48**(5):334–342. PMID: 21378379. doi: 10.1136/jmg.2010.087106.
- Blake, K.D., and Prasad, C. 2006. CHARGE syndrome. *Orphanet. J. Rare. Dis.* **1**:34. PMID: 16959034. doi: 10.1186/1750-1172-1-34.
- Boerkoel, C.F., Nowaczyk, M.J.M., Blaser, S.I., Meschino, W.S., and Weksberg, R. 1998. Schimke immunoosseous dysplasia complicated by moyamoya phenomenon. *Am. J. Med. Genet.* **78**(2):118–122. PMID: 9674900. doi: 10.1002/(SICI)1096-8628(19980630)78:2<118::AID-AJMG4>3.0.CO;2-K.
- Boerkoel, C.F., O'Neill, S., Andr, J.L., Benke, P.J., Bogdanov, R., Bulla, M., Burguet, A., Cockfield, S., Cordeiro, I., Ehrich, J.H.H., FrNd, S., Geary, D.F., Ieshima, A., Illies, F., Joseph, M.W., Kaitila, I., Lama, G., Leheup, B., Ludman, M.D., Mcleod, D.R., Medeira, A., Milford, D.V., Rm, T., Renner-Primec, Z., Santava, A., Santos, H.G., Schmidt, B., Smith, G.C., Spranger, J., Zupancic, N., and Weksberg, R. 2000. Manifestations and treatment of Schimke immuno-osseous dysplasia: 14 new cases and a review of the literature. *Eur. J. Pediatr.* **159**(1–2):1–7. PMID: 10653321. doi: 10.1007/s004310050001.
- Boisson, B., Laplantine, E., Prando, C., Giliani, S., Israelsson, E., Xu, Z., Abhyankar, A., Israël, L., Trevejo-Nunez, G., Bogunovic, D., Cepika, A.-M., Macduff, D., Chrabieh, M., Hubeau, M., Bajolle, F., Debré, M., Mazzolari, E., Vairo, D., Agou, F., Virgin, H.W., Bossuyt, X., Rambaud, C., Facchetti, F., Bonnet, D., Quartier, P., Fournet, J.-C., Pascual, V., Chaussabel, D., Notarangelo, L.D., Puel, A., Israël, A., Casanova, J.-L., and Picard, C. 2012. Immunodeficiency, autoinflammation and amylopectinosis in humans with inherited HOIL-1 and LUBAC deficiency. *Nat. Immunol.* **13**(12):1178–1186. PMID: 23104095. doi: 10.1038/ni.2457.
- Bolze, A., Byun, M., McDonald, D., Morgan, N.V., Abhyankar, A., Premkumar, L., Puel, A., Bacon, C. M., Rieux-Laucat, F., Pang, K., Britland, A., Abel, L., Cant, A., Maher, E.R., Riedl, S.J., Hambleton, S., and Casanova, J.-L. 2010. Whole-exome-sequencing-based discovery of human FADD deficiency. *Am. J. Hum. Genet.* **87**(6):873–881. PMID: 21109225. doi: 10.1016/j.ajhg.2010.10.028.
- Booth, C., Gilmour, K.C., Veys, P., Gennery, A.R., Slat-ter, M.A., Chapel, H., Heath, P.T., Steward, C.G., Smith, O., O'meara, A., Kerrigan, H., Mahlaoui, N., Cavazzana-Calvo, M., Fischer, A., Moshous, D., Blanche, S., Pachlopnik Schmid, J., Latour, S., De Saint-Basile, G., Albert, M., Notheis, G., Rieber, N., Strahm, B., Ritterbusch, H., Lankester, A., Hartwig, N.G., Meyts, I., Plebani, A., Soresina, A., Finocchi, A., Pignata, C., Cirillo, E., Bonanomi, S., Peters, C., Kalwak, K., Pasic, S., Sedlacek, P., Jazbec, J., Kanegane, H., Nichols, K.E., Hanson, I.C., Kapoor, N., Haddad, E., Cowan, M., Choo, S., Smart, J., Arkwright, P.D., and Gaspar, H. B. 2011. X-linked lymphoproliferative disease due to SAP/SH2D1A deficiency: a multicenter study on the manifestations, management and outcome of the disease. *Blood.* **117**(1):53–62. PMID: 20926771. doi: 10.1182/blood-2010-06-284935.
- Bouma, G., Burns, S.O., and Thrasher, A.J. 2009. Wiskott-Aldrich Syndrome: immunodeficiency resulting from defective cell migration and impaired immunostimulatory activation. *Immunobiology.* **214**(9–10): 778–790. PMID: 19628299. doi: 10.1016/j.imbio.2009.06.009.
- Bousfiha, A.A., Jeddane, L., Ailal, F., Al Herz, W., Con-ley, M.E., Cunningham-Rundles, C., Etzioni, A., Fischer, A., Franco, J.L., Geha, R.S., Hammarström, L., Nonoyama, S., Ochs, H. D., Roifman, C.M., Seger, R., Tang, M.L.K., Puck, J.M., Chapel, H., Notarangelo, L.D., and Casanova, J.-L. 2013. A phenotypic approach for IUIS PID classification and diagnosis guidelines for clinicians at the bedside. *J. Clin. Immunol.* **33**(6):1078–1087. PMID: 23657403. doi: 10.1007/s10875-013-9901-6.
- Boztug, K., Appaswamy, G., Ashikov, A., Schäffer, A.A., Salzer, U., Diestelhorst, J., Germeshausen, M., Brandes, G., Lee-Gossler, J., Noyan, F., Gatzke, A.-K., Minkov, M., Greil, J., Kratz, C., Petropoulou, T., Pellier, I., Bellanné-Chantelot, C., Rezaei, N., Mönke-möller, K., Irani-Hakimeh, N., Bakker, H., Gerardy-Schahn, R., Zeidler, C., Grimbacher, B., Welte, K., and Klein, C. 2009. A syndrome with congenital neutropenia and mutations in G6PC3. *N. Engl. J. Med.* **360**(1):32–43. PMID: 19118303. doi: 10.1056/NEJMoa0805051.
- Bruckmann, C., Lindner, W., Roos, R., Permanetter, W., Haas, R.J., Haworth, S.G., and Belohradsky, B.H. 1991. Severe pulmonary vascular occlusive disease following bone marrow transplantation in Omenn syndrome. *Eur. J. Pediatr.* **150**(4):242–245. PMID: 2029913. doi: 10.1007/BF01955521.
- Burroughs, L., Woolfrey, A., and Shimamura, A. 2009. Shwachman-Diamond syndrome: a review of the clinical presentation, molecular pathogenesis, diagnosis, and treatment. *Hematol. Oncol. Clin.*

- North Am. **23**(2):233–248. PMID: 19327581. doi: 10.1016/j.hoc.2009.01.007.
- Cavadini, P., Vermi, W., Facchetti, F., Fontana, S., Nagafuchi, S., Mazzolari, E., Sediva, A., Marrella, V., Villa, A., Fischer, A., Notarangelo, L.D., and Badolato, R. 2005. AIRE deficiency in thymus of 2 patients with Omenn syndrome. *J. Clin. Invest.* **115**(3):728–732. PMID: 15696198. doi: 10.1172/JCI23087; doi: 10.1172/JCI200523087.
- Chrzanowska, K.H., Gregorek, H., Dembowska-Baginska, B., Kalina, M.A., and Digweed, M. 2012. Nijmegen breakage syndrome (NBS). *Orphanet. J. Rare. Dis.* **7**:13. PMID: 22373003. doi: 10.1186/1750-1172-7-13.
- Clewing, J.M., Antalfy, B.C., Lucke, T., Najafian, B., Marwedel, K.M., Hori, A., Powel, R. M., Safo Do, A. F., Najera, L., Santacruz, K., John Hicks, M., Armstrong, D.L., and Boerkoel, C.F. 2007. Schimke immuno-osseous dysplasia: a clinicopathological correlation. *J. Med. Genet.* **44**(2):122–130. PMID: 16840568. doi: 10.1136/jmg.2006.044313.
- Conley, M.E., Dobbs, A.K., Farmer, D.M., Kilic, S., Paris, K., Grigoriadou, S., Coustan-Smith, E., Howard, V., and Campana, D. 2009. Primary B cell immunodeficiencies: comparisons and contrasts. *Annu. Rev. Immunol.* **27**:199–227. PMID: 19302039. doi: 10.1146/annurev.immunol.021908.132649.
- Crow, Y.J., Vanderver, A., Orcesi, S., Kuijpers, T.W., and Rice, G.I. 2014. Therapies in Aicardi-Goutieres syndrome. *Clin. Exp. Immunol.* **175**(1):1–8. PMID: 23607857. doi: 10.1111/cei.12115.
- Cunningham-Rundles, C. 2012. The many faces of common variable immunodeficiency. *Hematol. Am. Soc. Hematol. Educ. Prog.* **2012**:301–305. PMID: 23233596.
- Dickinson, R.E., Griffin, H., Bigley, V., Reynard, L.N., Hussain, R., Haniffa, M., Lakey, J. H., Rahman, T., Wang, X.-N., McGovern, N., Pagan, S., Cookson, S., McDonald, D., Chua, I., Wallis, J., Cant, A., Wright, M., Keavney, B., Chinnery, P.F., Loughlin, J., Hambleton, S., Santibanez-Koref, M., and Collin, M. 2011. Exome sequencing identifies GATA-2 mutation as the cause of dendritic cell, monocyte, B and NK lymphoid deficiency. *Blood.* **118**(10):2656–2658. PMID: 21765025. doi: 10.1182/blood-2011-06-360313.
- Dror, Y., Ginzberg, H., Dalal, I., Cherepanov, V., Downey, G., Durie, P., Roifman, C.M., and Freedman, M.H. 2001. Immune function in patients with Shwachman-Diamond syndrome. *Br. J. Haematol.* **114**(3):712–717. PMID: 11553003. doi: 10.1046/j.1365-2141.2001.02996.x.
- Dror, Y., Donadieu, J., Koglmeier, J., Dodge, J., Toivainen-Salo, S., Makitie, O., Kerr, E., Zeidler, C., Shimamura, A., Shah, N., Cipolli, M., Kuijpers, T., Durie, P., Rommens, J., Siderius, L., and Liu, J.M. 2011. Draft consensus guidelines for diagnosis and treatment of Shwachman-Diamond syndrome. *Ann. NY Acad. Sci.* **1242**:40–55. PMID: 22191555. doi: 10.1111/j.1749-6632.2011.06349.x.
- Dutz, J.P., Benoit, L., Wang, X., Demetrick, D.J., Junker, A., de Sa, D., and Tan, R. 2001. Lymphocytic vasculitis in X-linked lymphoproliferative disease. *Blood.* **97**(1):95–100. doi: 10.1182/blood.V97.1.95.
- Ege, M., Ma, Y., Manfras, B., Kalwak, K., Lu, H., Lieber, M.R., Schwarz, K., and Pannicke, U. 2005. Omenn syndrome due to ARTEMIS mutations. *Blood.* **105**(11):4179–4186. PMID: 15731174. doi: 10.1182/blood-2004-12-4861.
- Feske, S., Picard, C., and Fischer, A. 2010. Immunodeficiency due to mutations in ORAI1 and STIM1. *Clin. Immunol.* **135**(2):169–182. PMID: 20189884. doi: 10.1016/j.clim.2010.01.011.
- Finsterer, J., and Frank, M. 2013. Haematological features in Barth syndrome. *Curr. Opin. Hematol.* **20**(1):36–40. PMID: 23041719. doi: 10.1097/MOH.0b013e32835a01d9.
- Ganaie, H., Eisenstein, E.M., Tenenbaum, A., Somech, R., Simanovsky, N., Roscioli, T., Weintraub, M., and Stepensky, P. 2013. The role of hematopoietic stem cell transplantation in SP110 associated veno-occlusive disease with immunodeficiency syndrome. *Pediatr. Allergy. Immunol.* **24**(3):250–256. PMID: 23448538. doi: 10.1111/pai.12051.
- Gennery, A.R. 2012. Immunological aspects of 22q11.2 deletion syndrome. *Cell. Mol. Life. Sci.* **69**(1):17–27. PMID: 21984609. doi: 10.1007/s00018-011-0842-z.
- Gennery, A.R., Slatter, M.A., Rice, J., Hoefsloot, L.H., Barge, D., Mclean-Tooke, A., Montgomery, T., Goodship, J.A., Burt, A.D., Flood, T.J., Abinun, M., Cant, A.J., and Johnson, D. 2008. Mutations in CHD7 in patients with CHARGE syndrome cause T-B + natural killer cell + severe combined immune deficiency and may cause Omenn-like syndrome. *Clin. Exp. Immunol.* **153**(1):75–80. PMID: 18505430. doi: 10.1111/j.1365-2249.2008.03681.x.
- Grouhi, M., Dalal, I., Nisbet-Brown, E., and Roifman, C. M. 1998. Cerebral vasculitis associated with chronic mucocutaneous candidiasis. *J. Pediatr.* **133**(4):571–574. doi: 10.1016/S0022-3476(98)70072-1.

- Grunebaum, E., Bates, A., and Roifman, C.M. 2008. Omenn syndrome is associated with mutations in DNA ligase IV. *J. Allergy. Clin. Immunol.* **122** (6):1219–1220. PMID: 18845326. doi: 10.1016/j.jaci.2008.08.031.
- Hauck, F., Randriamampita, C., Martin, E., Gerart, S., Lambert, N., Lim, A., Soulier, J., Maciorowski, Z., Touzot, F., Moshous, D., Quartier, P., Heritier, S., Blanche, S., Rieux-Laucat, F., Brousse, N., Callebaut, I., Veillette, A., Hivroz, C., Fischer, A., Latour, S., and Picard, C. 2012. Primary T-cell immunodeficiency with immunodysregulation caused by autosomal recessive LCK deficiency. *J. Allergy. Clin. Immunol.* **130**(5):1144–1152 e11. PMID: 22985903. doi: 10.1016/j.jaci.2012.07.029.
- Heimall, J., Freeman, A., and Holland, S.M. 2010. Pathogenesis of hyper IgE syndrome. *Clin. Rev. Allergy. Immunol.* **38**(1):32–38. PMID: 19452285. doi: 10.1007/s12016-009-8134-1.
- Hoernes, M., Seger, R., and Reichenbach, J. 2011. Modern management of primary B-cell immunodeficiencies. *Pediatr. Allergy. Immunol.* **22**(8):758–769. PMID: 22122788. doi: 10.1111/j.1399-3038.2011.01236.x.
- Hori, T., Ohnishi, H., Teramoto, T., Tsubouchi, K., Naiki, T., Hirose, Y., Ohara, O., Seishima, M., Kaneko, H., Fukao, T., and Kondo, N. 2012. Autosomal-dominant chronic mucocutaneous candidiasis with STAT1-mutation can be complicated with chronic active hepatitis and hypothyroidism. *J. Clin. Immunol.* **32**(6):1213–1220. PMID: 22847544. doi: 10.1007/s10875-012-9744-6.
- Hsu, A.P., Sampaio, E.P., Khan, J., Calvo, K.R., Lemieux, J.E., Patel, S.Y., Frucht, D.M., Vinh, D.C., Auth, R.D., Freeman, A.F., Olivier, K.N., Uzel, G., Zerbe, C.S., Spalding, C., Pittaluga, S., Raffeld, M., Kuhns, D.B., Ding, L., Paulson, M.L., Marciano, B.E., Gea-Banacloche, J.C., Orange, J.S., Cuellar-Rodriguez, J., Hickstein, D.D., and Holland, S.M. 2011. Mutations in GATA2 are associated with the autosomal dominant and sporadic monocytopenia and mycobacterial infection (MonoMAC) syndrome. *Blood.* **118**(10):2653–2655. PMID: 21670465. doi: 10.1182/blood-2011-05-356352.
- Hsu, A.P., Sowerwine, K.J., Lawrence, M.G., Davis, J., Henderson, C.J., Zarembek, K.A., Garofalo, M., Gallin, J.I., Kuhns, D.B., Heller, T., Milner, J.D., Puck, J.M., Freeman, A.F., and Holland, S.M. 2013. Intermediate phenotypes in patients with autosomal dominant hyper-IgE syndrome caused by somatic mosaicism. *J. Allergy. Clin. Immunol.* **131**(6):1586–1593. PMID: 23623265. doi: 10.1016/j.jaci.2013.02.038.
- Jain, M., Jakubowski, A., Cui, L., Shi, J., Su, L., Bauer, M., Guan, J., Lim, C.C., Naito, Y., Thompson, J.S., Sam, F., Ambrose, C., Parr, M., Crowell, T., Lincecum, J.M., Wang, M.Z., Hsu, Y.-M., Zheng, T.S., Michaelson, J.S., Liao, R., and Burkly, L.C. 2009. A novel role for tumor necrosis factor-like weak inducer of apoptosis (TWEAK) in the development of cardiac dysfunction and failure. *Circulation.* **119** (15):2058–2068. PMID: 19349318. doi: 10.1161/CIRCULATIONAHA.108.837286.
- Jefferies, J.T.C., Chin, A.J., and Jefferies, J.L. 2013. Barth syndrome. *Am. J. Med. Genet. C. Semin. Med. Genet.* **163C**(3):198–205. PMID: 23843353. doi: 10.1002/ajmg.c.31372.
- Johnson, K.D., Hsu, A.P., Ryu, M.-J., Wang, J., Gao, X., Boyer, M.E., Liu, Y., Lee, Y., Calvo, K.R., Keles, S., Zhang, J., Holland, S.M., and Bresnick, E.H. 2012. Cis-element mutated in GATA2-dependent immunodeficiency governs hematopoiesis and vascular integrity. *J. Clin. Invest.* **122**(10):3692–3704. PMID: 22996659. doi: 10.1172/JCI61623.
- Johnston, S.L., Hill, S.J., Lock, R.J., Dwight, J.F., Unsworth, D.J., and Gompels, M. M. 2004. Echocardiographic abnormalities in primary antibody deficiency. *Postgrad. Med. J.* **80**(942):214–218. PMID: 15082842. doi: 10.1136/pgmj.2003.012443.
- Jones, L.B.K.R., McGrogan, P., Flood, T.J., Gennery, A. R., Morton, L., Thrasher, A., Goldblatt, D., Parker, L., and Cant, A.J. 2008. Special article: chronic granulomatous disease in the United Kingdom and Ireland: a comprehensive national patient-based registry. *Clin. Exp. Immunol.* **152**(2):211–218. PMID: 18410635. doi: 10.1111/j.1365-2249.2008.03644.x.
- Jyonouchi, S., McDonald-McGinn, D.M., Bale, S., Zackai, E.H., and Sullivan, K.E. 2009. CHARGE (coloboma, heart defect, atresia choanae, retarded growth and development, genital hypoplasia, ear anomalies/deafness) syndrome and chromosome 22q11.2 deletion syndrome: a comparison of immunologic and nonimmunologic phenotypic features. *Pediatrics.* **123**(5):e871–e877. PMID: 19403480. doi: 10.1542/peds.2008-3400.
- Kamae, C., Nakagawa, N., Sato, H., Honma, K., Mitsuiki, N., Ohara, O., Kanegane, H., Pasic, S., Pan-Hammarström, Q., Van Zelm, M.C., Morio, T., Imai, K., and Nonoyama, S. 2013. Common variable immunodeficiency classification by quantifying T-cell receptor and immunoglobulin kappa-deleting recombination excision circles. *J. Allergy. Clin.*

- Immunol. **131**(5):1437–1440 e5. PMID: 23273952. doi: 10.1016/j.jaci.2012.10.059.
- Kashef, S., Moghtaderi, M., Borzouee, M., and Rezaei, N. 2011. Giant aneurysm of thoracic and proximal abdominal aorta in a patient with common variable immunodeficiency. *Iran. J. Allergy. Asthma. Immunol.* **10**(2):133–137. PMID: 21625023. doi: 10.02/ijaai.133137.
- Kavanagh, D., Spitzer, D., Kothari, P., Shaikh, A., Liszewski, M.K., Richards, A., and Atkinson, J.P. 2008. New roles for the major human 3'-5' exonuclease TREX1 in human disease. *Cell Cycle.* **7**(12):1718–1725. PMID: 18583934. doi: 10.4161/cc.7.12.6162.
- Kawai, T., and Malech, H.L. 2009. WHIM syndrome: congenital immune deficiency disease. *Curr. Opin. Hematol.* **16**(1):20–26. PMID: 19057201. doi: 10.1097/MOH.0b013e32831ac557.
- Kazenwadel, J., Secker, G.A., Liu, Y.J., Rosenfeld, J.A., Wildin, R.S., Cuellar-Rodriguez, J., Hsu, A.P., Dyack, S., Fernandez, C.V., Chong, C.-E., Babic, M., Bardy, P.G., Shimamura, A., Zhang, M.Y., Walsh, T., Holland, S.M., Hickstein, D.D., Horwitz, M.S., Hahn, C. N., Scott, H.S., and Harvey, N.L. 2012. Loss-of-function germline GATA2 mutations in patients with MDS/AML or MonoMAC syndrome and primary lymphedema reveal a key role for GATA2 in the lymphatic vasculature. *Blood.* **119**(5):1283–1291. PMID: 22147895. doi: 10.1182/blood-2011-08-374363.
- Kirkpatrick, C.H. 2001. Chronic mucocutaneous candidiasis. *Pediatr. Infect. Dis. J.* **20**(2):197–206. PMID: 11224843. doi: 10.1097/00006454-200102000-00017.
- Kivitie-Kallio, S., and Norio, R. 2001. Cohen syndrome: essential features, natural history, and heterogeneity. *Am. J. Med. Genet.* **102**(2):125–135. PMID: 11477603. doi: 10.1002/1096-8628(20010801)102:2<125::AID-AJMG1439>3.0.CO;2-0.
- Kivitie-Kallio, S., Eronen, M., Lipsanen-Nyman, M., Marttinen, E., and Norio, R. 1999. Cohen syndrome: evaluation of its cardiac, endocrine and radiological features. *Clin. Genet.* **56**(1):41–49. PMID: 10466416. doi: 10.1034/j.1399-0004.1999.560106.x.
- Kobrynski, L.J., and Sullivan, K.E. 2007. Velocardiofacial syndrome, DiGeorge syndrome: the chromosome 22q11.2 deletion syndromes. *Lancet.* **370**(9596):1443–1452. PMID: 17950858. doi: 10.1016/S0140-6736(07)61601-8.
- Kolehmainen, J., Black, G.C.M., Saarinen, A., Chandler, K., Clayton-Smith, J., Träskelin, A.-L., Perveen, R., Kivitie-Kallio, S., Norio, R., Warburg, M., Fryns, J.-P., Chapelle, A.D.L., and Lehesjoki, A.-E. 2003. Cohen syndrome is caused by mutations in a novel gene, COH1, encoding a transmembrane protein with a presumed role in vesicle-mediated sorting and intracellular protein transport. *Am. J. Hum. Genet.* **72**(6):1359–1369. PMID: 12730828. doi: 10.1086/375454.
- Kopel, L., Gutierrez, P.S., and Lage, S.G. 2011. Dilated cardiomyopathy in a case of Shwachman–Diamond syndrome. *Cardiol. Young.* **21**(5):588–590. PMID: 21486516. doi: 10.1017/S1047951111000308.
- Laufs, H., Nigrovic, P.A., Schneider, L.C., Oettgen, H., Del Nido, P., Moskowitz, I.P.G., Blume, E., and Perez-Atayde, A.R. 2002. Giant cell myocarditis in a 12-year-old girl with common variable immunodeficiency. *Mayo. Clin. Proc.* **77**(1):92–96. PMID: 11795251. doi: 10.4065/77.1.92.
- Leiding, J.W., Marciano, B.E., Zerbe, C.S., Deravin, S.S., Malech, H.L., and Holland, S.M. 2013. Diabetes, renal and cardiovascular disease in p47 phox-/- chronic granulomatous disease. *J. Clin. Immunol.* **33**(4):725–730. PMID: 23386289. doi: 10.1007/s10875-013-9871-8.
- Lev, A., Amariglio, N., Levy, Y., Spierer, Z., Anikster, Y., Rechavi, G., Dekel, B., and Somech, R. 2009. Molecular assessment of thymic capacities in patients with Schimke immuno-osseous dysplasia. *Clin. Immunol.* **133**(3):375–381. PMID: 19796992. doi: 10.1016/j.clim.2009.08.017.
- Liu, L., Okada, S., Kong, X.-F., Kreins, A.Y., Cypowyj, S., Abhyankar, A., Toubiana, J., Itan, Y., Audry, M., Nitschke, P., Masson, C., Toth, B., Flatot, J., Migaud, M., Chrabieh, M., Kochetkov, T., Bolze, A., Borghesi, A., Toulon, A., Hiller, J., Eyerich, S., Eyerich, K., Gulacsy, V., Chernyshova, L., Chernyshov, V., Bondarenko, A., Maria Cortes Grimaldo, R., Blancas-Galicia, L., Madrigal Beas, I.M., Roesler, J., Magdorf, K., Engelhard, D., Thumerelle, C., Burgel, P.-R., Hoernes, M., Drexel, B., Seger, R., Kusuma, T., Jansson, A.F., Sawalle-Belohradsky, J., Belohradsky, B., Jouanguy, E., Bustamante, J., Bue, M., Karin, N., Wildbaum, G., Bodemer, C., Lortholary, O., Fischer, A., Blanche, S., Al-Muhsen, S., Reichenbach, J., Kobayashi, M., Rosales, F.E., Lozano, C.T., Kilic, S.S., Oleastro, M., Etzioni, A., Traidl-Hoffmann, C., Renner, E.D., Abel, L., Picard, C., Marodi, L., Boisson-Dupuis, S., Puel, A., and Casanova, J.-L. 2011. Gain-of-function human STAT1 mutations impair IL-17 immunity and underlie chronic mucocutaneous candidiasis. *J. Exp. Med.* **208**(8):1635–1648. PMID: 21727188. doi: 10.1084/jem.20110958.
- Loffredo, L., Carnevale, R., Sanguigni, V., Plebani, A., Rossi, P., Pignata, C., De Mattia, D., Finocchi, A.,

- Martire, B., Pietrogrande, M.C., Martino, S., Gambineri, E., Giardino, G., Soresina, A.R., Martino, F., Pignatelli, P., and Violi, F. 2013. Does NADPH oxidase deficiency cause artery dilatation in humans? *Antioxid. Redox. Signal.* **18**(12):1491–1496. PMID: 23216310. doi: 10.1089/ars.2012.4987.
- Mahlaoui, N., Pellier, I., Mignot, C., Jais, J.-P., Bilhou-Nabera, C., Moshous, D., Neven, B., Picard, C., De Saint-Basile, G., Cavazzana-Calvo, M., Blanche, S., and Fischer, A. 2013. Characteristics and outcome of early-onset, severe forms of Wiskott-Aldrich syndrome. *Blood.* **121**(9):1510–1516. PMID: 23264593. doi: 10.1182/blood-2012-08-448118.
- Mandel, K., Grunebaum, E., and Benson, L. 2001. Non-compaction of the myocardium associated with Roifman syndrome. *Cardiol. Young.* **11**(2):240–243. PMID: 11293748. doi: 10.1017/S1047951101000208.
- Marazzi, M.G., Bondi, E., Giannattasio, A., Strozzi, M., and Savioli, C. 2008. Intracranial aneurysm associated with chronic mucocutaneous candidiasis. *Eur. J. Pediatr.* **167**(4):461–463. PMID: 17443345. doi: 10.1007/s00431-007-0490-3.
- Marciano, B.E., Rosenzweig, S.D., Kleiner, D.E., Anderson, V.L., Darnell, D.N., Anaya-O'Brien, S., Hilligoss, D.M., Malech, H.L., Gallin, J.I., and Holland, S.M. 2004. Gastrointestinal involvement in chronic granulomatous disease. *Pediatrics.* **114**(2):462–468. PMID: 15286231. doi: 10.1542/peds.114.2.462.
- McCarl, C.-A., Picard, C., Khalil, S., Kawasaki, T., Röther, J., Papolos, A., Kutok, J., Hivroz, C., Ledeist, F., Plogmann, K., Ehl, S., Notheis, G., Albert, M.H., Belohradsky, B.H., Kirschner, J., Rao, A., Fischer, A., and Feske, S. 2009. ORAI1 deficiency and lack of store-operated Ca²⁺ entry cause immunodeficiency, myopathy, and ectodermal dysplasia. *J. Allergy. Clin. Immunol.* **124**(6):1311–1318 e7. PMID: 20004786. doi: 10.1016/j.jaci.2009.10.007.
- McCluggage, W.G., Armstrong, D.J., Maxwell, R.J., Ellis, P.K., and McCluskey, D.R. 1999. Systemic vasculitis and aneurysm formation in the Wiskott-Aldrich syndrome. *J. Clin. Pathol.* **52**(5):390–392. PMID: 10560364. doi: 10.1136/jcp.52.5.390.
- McDermott, D.H., De Ravin, S.S., Jun, H.S., Liu, Q., Priel, D.A.L., Noel, P., Takemoto, C.M., Ojode, T., Paul, S.M., Dunsmore, K.P., Hilligoss, D., Marquesen, M., Ulrick, J., Kuhns, D.B., Chou, J.Y., Malech, H.L., and Murphy, P.M. 2010. Severe congenital neutropenia resulting from G6PC3 deficiency with increased neutrophil CXCR4 expression and myelokathexis. *Blood.* **116**(15):2793–2802. PMID: 20616219. doi: 10.1182/blood-2010-01-265942.
- Mehes, K., Kosztolányi, G., Kardos, M., Horváth, M., Opitz, J.M., and Reynolds, J.F. 1988. Cohen syndrome: a connective tissue disorder? *Am. J. Med. Genet.* **31**(1): 131–133. PMID: 3223494. doi: 10.1002/ajmg.1320310115.
- Momma, K. 2010. Cardiovascular anomalies associated with chromosome 22q11.2 deletion syndrome. *Am. J. Cardiol.* **105**(11):1617–1624. PMID: 20494672. doi: 10.1016/j.amjcard.2010.01.333.
- Morimoto, M., Yu, Z., Stenzel, P., Clewing, J., Najafian, B., Mayfield, C., Henderson, G., Weinkauff, J.G., Gormley, A.K., Parham, D.M., Ponniah, U., André, J.-L., Asakura, Y., Basiratnia, M., Bogdanovic, R., Bokenkamp, A., Bonneau, D., Buck, A., Charrow, J., Cochat, P., Cordeiro, I., Deschenes, G., Fenkçi, M., Frange, P., Fründ, S., Fryssira, H., Guillen-Navarro, E., Keller, K., Kirmani, S., Kobelka, C., Lamfers, P., Levchenko, E., Lewis, D.B., Massella, L., Mcleod, D., Milford, D.V., Nobili, F., Saraiva, J.M., Semerci, C., Shoemaker, L., Stajic, N., Stein, A., Taha, D., Wand, D., Zonana, J., Lücke, T., and Boerkoel, C.F. 2012. Reduced elastogenesis: a clue to the arteriosclerosis and emphysematous changes in Schimke immuno-osseous dysplasia? *Orphanet. J. Rare. Dis.* **7**:70. PMID: 22998683. doi: 10.1186/1750-1172-7-70.
- Myers, K.C., Bolyard, A.A., Otto, B., Wong, T.E., Jones, A.T., Harris, R.E., Davies, S.M., Dale, D.C., and Shimamura, A. 2014. Variable clinical presentation of Shwachman–Diamond syndrome: update from the North American Shwachman–Diamond Syndrome Registry. *J. Pediatr.* **164**(4):866–870. PMID: 24388329. doi: 10.1016/j.jpeds.2013.11.039.
- Nagashima, T., Miyanooshita, A., Sakiyama, Y., Ozaki, Y., Stan, A.C., and Nagashima, K. 2000. Cerebral vasculitis in chronic mucocutaneous candidiasis: autopsy case report. *Neuropathology.* **20**(4):309–314. PMID: 11211056. doi: 10.1111/j.1440-1789.2000.00352.x; doi: 10.1046/j.1440-1789.2000.00352.x.
- Nahum, A., Reid, B., Grunebaum, E., and Roifman, C. M. 2009. Matched unrelated bone marrow transplant for Omenn syndrome. *Immunol. Res.* **44**(1–3):25–34. PMID: 18854956. doi: 10.1007/s12026-008-8067-4.
- Nahum, A., Dadi, H., Bates, A., and Roifman, C.M. 2012. The biological significance of TLR3 variant, L412F, in conferring susceptibility to cutaneous candidiasis, CMV and autoimmunity. *Autoimmun. Rev.* **11**(5):341–347. PMID: 22024499. doi: 10.1016/j.autrev.2011.10.007.
- Nanetti, L., Cavalieri, S., Pensato, V., Erbetta, A., Pareyson, D., Panzeri, M., Zorzi, G., Antozzi, C., Moroni, I., Gellera, C., Brusco, A., and Mariotti, C. 2013. SETX

- mutations are a frequent genetic cause of juvenile and adult onset cerebellar ataxia with neuropathy and elevated serum alpha-fetoprotein. *Orphanet. J. Rare. Dis.* **8**:123. PMID: 23941260. doi: 10.1186/1750-1172-8-123.
- Nehme, N.T., Schmid, J.P., Debeurme, F., Andre-Schmutz, I., Lim, A., Nitschke, P., Rieux-Laucat, F., Lutz, P., Picard, C., Mahlaoui, N., Fischer, A., and De Saint Basile, G. 2012. MST1 mutations in autosomal recessive primary immunodeficiency characterized by defective naive T-cell survival. *Blood*. **119** (15):3458–3468. PMID: 22174160. doi: 10.1182/blood-2011-09-378364.
- Notarangelo, L.D. 2013. Partial defects of T-cell development associated with poor T-cell function. *J. Allergy. Clin. Immunol.* **131**(5):1297–1305. PMID: 23465662. doi: 10.1016/j.jaci.2013.01.020.
- Ochs, H.D., Filipovich, A.H., Veys, P., Cowan, M.J., and Kapoor, N. 2009. Wiskott-Aldrich syndrome: diagnosis, clinical and laboratory manifestations, and treatment. *Biol. Blood. Marrow. Transpl.* **15**(1 Suppl):84–90. PMID: 19147084. doi: 10.1016/j.bbmt.2008.10.007.
- Ono, M., Goerler, H., and Breymann, T. 2009. Aneurysm of the aortic root in the setting of Wiskott-Aldrich syndrome. *Cardiol. Young.* **19**(2):212–215. PMID: 19195418. doi: 10.1017/S1047951109003564.
- Ostergaard, P., Simpson, M.A., Connell, F.C., Steward, C.G., Brice, G., Woollard, W.J., Dafou, D., Kilo, T., Smithson, S., Lunt, P., Murday, V.A., Hodgson, S., Keenan, R., Pilz, D.T., Martinez-Corral, I., Makinen, T., Mortimer, P.S., Jeffery, S., Trembath, R.C., and Mansour, S. 2011. Mutations in GATA2 cause primary lymphedema associated with a predisposition to acute myeloid leukemia (Emberger syndrome). *Nat. Genet.* **43**(10):929–931. PMID: 21892158. doi: 10.1038/ng.923.
- Pachlopnik Schmid, J. 2012. Polymerase epsilon1 mutation in a human syndrome with facial dysmorphism, immunodeficiency, livedo, and short stature (“FILS syndrome”). *J. Exp. Med.* **209**(13):2323–2330. PMID: 23230001.
- Pellier, I., Girod, S.D., Loisel, D., Benabidallah, S., Proust, A., Malhlaoui, N., Picard, C., Najioullah, F., De Saint Basile, G., Blanche, S., Rialland, X., Casanova, J.L., and Fischer, A. 2011. Occurrence of aortic aneurysms in 5 cases of Wiskott-Aldrich syndrome. *Pediatrics.* **127**(2):e498–e504. PMID: 21262885. doi: 10.1542/peds.2009-2987.
- Perlman, S.L., Boder (Deceased), E., Sedgewick, R.P., and Gatti, R.A. 2012. Ataxia-telangiectasia. *Handb. Clin. Neurol.* **103**:307–332. PMID: 21827897. doi: 10.1016/B978-0-444-51892-7.00019-X.
- Poliani, P.L., Facchetti, F., Ravanini, M., Gennery, A.R., Villa, A., Roifman, C.M., and Notarangelo, L.D. 2009. Early defects in human T-cell development severely affect distribution and maturation of thymic stromal cells: possible implications for the pathophysiology of Omenn syndrome. *Blood*. **114**(1):105–108. PMID: 19414857. doi: 10.1182/blood-2009-03-211029.
- Puel, A., Doffinger, R., Natividad, A., Chrabieh, M., Barchenas-Morales, G., Picard, C., Cobat, A., Ouachee-Chardin, M., Toulon, A., Bustamante, J., Al-Muhsen, S., Al-Owain, M., Arkwright, P.D., Costigan, C., McConnell, V., Cant, A.J., Abinun, M., Polak, M., Bougneres, P.-F., Kumararatne, D., Marodi, L., Nahum, A., Roifman, C., Blanche, S., Fischer, A., Bodemer, C., Abel, L., Lilic, D., and Casanova, J.-L. 2010. Autoantibodies against IL-17A, IL-17F, and IL-22 in patients with chronic mucocutaneous candidiasis and autoimmune polyendocrine syndrome type I. *J. Exp. Med.* **207**(2):291–297. PMID: 20123958. doi: 10.1084/jem.20091983.
- Roifman, C.M. 1999. Antibody deficiency, growth retardation, spondyloepiphyseal dysplasia and retinal dystrophy: a novel syndrome. *Clin. Genet.* **55**(2):103–109. PMID: 10189087. doi: 10.1034/j.1399-0004.1999.550206.x.
- Roifman, C.M., Somech, R., Kavadas, F., Pires, L., Nahum, A., Dalal, I., and Grunebaum, E. 2012. Defining combined immunodeficiency. *J. Allergy. Clin. Immunol.* **130**(1):177–183. PMID: 22664165. doi: 10.1016/j.jaci.2012.04.029.
- Roscioli, T., Cliffe, S.T., Bloch, D.B., Bell, C.G., Mullan, G., Taylor, P.J., Sarris, M., Wang, J., Donald, J.A., Kirk, E.P., Ziegler, J.B., Salzer, U., McDonald, G.B., Wong, M., Lindeman, R., and Buckley, M.F. 2006. Mutations in the gene encoding the PML nuclear body protein Sp110 are associated with immunodeficiency and hepatic veno-occlusive disease. *Nat. Genet.* **38**(6):620–622. PMID: 16648851. doi: 10.1038/ng1780.
- Sack, J., and Friedman, E. 1980. Cardiac involvement in the Cohen syndrome: a case report. *Clin. Genet.* **17** (5):317–319. doi: 10.1111/j.1399-0004.1980.tb00156.x.
- Sampaio, E.P., Hsu, A.P., Pechacek, J., Bax, H.I., Dias, D. L., Paulson, M.L., Chandrasekaran, P., Rosen, L.B., Carvalho, D.S., Ding, L., Vinh, D.C., Browne, S.K., Datta, S., Milner, J.D., Kuhns, D.B., Long Priel, D.A., Sadat, M.A., Shiloh, M., De Marco, B., Alvares, M., Gillman, J.W., Ramarathnam, V., De La Morena, M.,

- Bezrodnik, L., Moreira, I., Uzel, G., Johnson, D., Spalding, C., Zerbe, C.S., Wiley, H., Greenberg, D.E., Hoover, S.E., Rosenzweig, S.D., Galgiani, J.N., and Holland, S.M. 2013. Signal transducer and activator of transcription 1 (STAT1) gain-of-function mutations and disseminated coccidioidomycosis and histoplasmosis. *J. Allergy. Clin. Immunol.* **131**(6):1624–1634. PMID: 23541320. doi: 10.1016/j.jaci.2013.01.052.
- Schlame, M., and Ren, M. 2006. Barth syndrome, a human disorder of cardiolipin metabolism. *FEBS Lett.* **580**(23):5450–5455. PMID: 16973164. doi: 10.1016/j.febslet.2006.07.022.
- Sen, S., Wang, H., Nghiem, C.L., Zhou, K., Yau, J., Tailor, C.S., Irwin, M.S., and Dror, Y. 2011. The ribosome-related protein, SBDS, is critical for normal erythropoiesis. *Blood.* **118**(24):6407–6417. PMID: 21963601. doi: 10.1182/blood-2011-02-335190.
- Somech, R., Simon, A.J., Lev, A., Dalal, I., Spierer, Z., Goldstein, I., Nagar, M., Amariglio, N., Rechavi, G., and Roifman, C.M. 2009. Reduced central tolerance in Omenn syndrome leads to immature self-reactive oligoclonal T cells. *J. Allergy. Clin. Immunol.* **124**(4):793–800. PMID: 19767069. doi: 10.1016/j.jaci.2009.06.048.
- Spencer, C.T., Bryant, R.M., Day, J., Gonzalez, I.L., Colan, S.D., Thompson, W.R., Berthy, J., Redfearn, S.P., and Byrne, B.J. 2006. Cardiac and clinical phenotype in Barth syndrome. *Pediatrics.* **118**(2):e337–e346. PMID: 16847078. doi: 10.1542/peds.2005-2667.
- Su, Y., and Swift, M. 2000. Mortality rates among carriers of ataxia-telangiectasia mutant alleles. *Ann. Intern. Med.* **133**(10):770–778. PMID: 11085839. doi: 10.7326/0003-4819-133-10-200011210-00009.
- Tekin, M., Dogu, F., Taçyildiz, N., Akar, E., İkinciogullari, A., Ogur, G., Yavuz, G., Babacan, E., and Akar, N. 2002. 657del5 mutation in the NBS1 gene is associated with Nijmegen breakage syndrome in a Turkish family. *Clin. Genet.* **62**(1):84–88. PMID: 12123493. doi: 10.1034/j.1399-0004.2002.620112.x.
- Toiviainen-Salo, S., Pitkänen, O., Holmström, M., Koikkalainen, J., Lötjönen, J., Lauerma, K., Taskinen, M., Savilahti, E., Smallhorn, J., Mäkitie, O., and Kivistö, S. 2008. Myocardial function in patients with Shwachman-Diamond syndrome: aspects to consider before stem cell transplantation. *Pediatr. Blood. Cancer.* **51**(4):461–467. PMID: 18646182. doi: 10.1002/pbc.21686.
- Uzel, G., Sampaio, E.P., Lawrence, M.G., Hsu, A.P., Hackett, M., Dorsey, M.J., Noel, R.J., Verbsky, J.W., Freeman, A.F., Janssen, E., Bonilla, F.A., Pechacek, J., Chandrasekaran, P., Browne, S.K., Agharahami, A., Gharib, A.M., Mannurita, S.C., Yim, J.J., Gambineri, E., Torgerson, T., Tran, D.Q., Milner, J.D., and Holland, S.M. 2013. Dominant gain-of-function STAT1 mutations in FOXP3 wild-type immune dysregulation–polyendocrinopathy–enteropathy–X-linked–like syndrome. *J. Allergy. Clin. Immunol.* **131**(6):1611–1623. PMID: 23534974. doi: 10.1016/j.jaci.2012.11.054.
- Villa, A., Notarangelo, L.D., and Roifman, C.M. 2008. Omenn syndrome: inflammation in leaky severe combined immunodeficiency. *J. Allergy. Clin. Immunol.* **122**(6):1082–1086. PMID: 18992930. doi: 10.1016/j.jaci.2008.09.037.
- Villa, A., Santagata, S., Bozzi, F., Giliani, S., Frattini, A., Imberti, L., Gatta, L.B., Ochs, H. D., Schwarz, K., Notarangelo, L.D., Vezzoni, P., and Spanopoulou, E. 1998. Partial V(D)J recombination activity leads to Omenn syndrome. *Cell.* **93**(5):885–896. PMID: 9630231. doi: 10.1016/S0092-8674(00)81448-8.
- Volkers, M., Dolatabadi, N., Gude, N., Most, P., Sussman, M.A., and Hassel, D. 2012. Orail deficiency leads to heart failure and skeletal myopathy in zebrafish. *J. Cell. Sci.* **125**(Pt 2):287–294. PMID: 22302996. doi: 10.1242/jcs.090464.
- Wang, H.-Y., Ma, C.A., Zhao, Y., Fan, X., Zhou, Q., Edmonds, P., Uzel, G., Oliveira, J.B., Orange, J., and Jain, A. 2013. Antibody deficiency associated with an inherited autosomal dominant mutation in TWEAK. *Proc. Natl. Acad. Sci. USA.* **110**(13):5127–5132. PMID: 23493554. doi: 10.1073/pnas.1221211110.
- Weemaes, C.M., Van Tol, M.J., Wang, J., Van Ostaijen-Ten Dam, M.M., Van Eggermond, M.C., Thijssen, P. E., Aytekin, C., Brunetti-Pierri, N., Van Der Burg, M., Graham Davies, E., Ferster, A., Furthner, D., Gimelli, G., Gennery, A., Kloeckener-Gruissem, B., Meyn, S., Powell, C., Reisli, I., Schuetz, C., Schulz, A., Shugar, A., Van Den Elsen, P.J., and Van Der Maarel, S.M. 2013. Heterogeneous clinical presentation in ICF syndrome: correlation with underlying gene defects. *Eur. J. Hum. Genet.* **21**(11):1219–1225. PMID: 23486536. doi: 10.1038/ejhg.2013.40.
- Wilda, M., Demuth, I., Concannon, P., Sperling, K., and Hameister, H. 2000. Expression pattern of the Nijmegen breakage syndrome gene, Nbs1, during murine development. *Human Molecular Genetics.* **9**(12):1739–1744. PMID: 3223494. doi: 10.1093/hmg/9.12.1739.
- Witzl, K., Cale, C.M., Pierce, C.M., Wilson, L.C., and Hennekam, R.C.M. 2007. Immunological abnormalities in CHARGE syndrome. *Eur. J. Med. Genet.*

- 50**(5):338–345. PMID: 17684005. doi: 10.1016/j.ejmg.2007.05.002.
- Yalcin, A., Aydemir, N., Turgut, H., Erbay, R., and Yalcin, A. 2004. Common variable immunodeficiency syndrome with right aortic arch: a case report. *BMC Infect. Dis.* **4**:2. PMID: 15005811. doi: 10.1186/1471-2334-4-2.
- Yamamoto, S., Yang, G., Zablocki, D., Liu, J., Hong, C., Kim, S.-J., Soler, S., Odashima, M., Thaisz, J., Yehia, G., Molina, C.A., Yatani, A., Vatner, D.E., Vatner, S. F., and Sadoshima, J. 2003. Activation of Mst1 causes dilated cardiomyopathy by stimulating apoptosis without compensatory ventricular myocyte hypertrophy. *J. Clin. Invest.* **111**(10):1463–1474. PMID: 12750396. doi: 10.1172/JCI200317459; doi: 10.1172/JCI17459.
- Yavuz, H., and Chee, R. 2010. A review on the vascular features of the hyperimmunoglobulin E syndrome. *Clin. Exp. Immunol.* **159**(3):238–244. PMID: 19912258. doi: 10.1111/j.1365-2249.2009.04044.x.
- Zhang, J., Quintal, L., Atkinson, A., Williams, B., Grunebaum, E., and Roifman, C.M. 2005. Novel RAG1 mutation in a case of severe combined immunodeficiency. *Pediatrics.* **116**(3):e445–e449. doi: 10.1542/peds.2005-0369.

Appendices

The appendices begin on the following page.

Appendix A1: Cardiac and vascular abnormalities associated with primary immunodeficiency disorders.

PID	Vascular abnormalities	Cardiac abnormalities
Predominantly T-cell lineage		
Omenn Syndrome	—	Eosinophil-mediated cardiac cytotoxicity and endomyocardial disease leading to cardiomyopathy
Calcium channel deficiency: <i>ORAI1</i> deficiency	—	Cardiomyopathy, bradycardia, heart failure
AD chronic mucocutaneous candidiasis: <i>STAT1</i> deficiency	Cerebral vasculitis, intracranial aneurysms and secondary infarctions	
<i>STK4</i> deficiency		ASD, mitral/tricuspid/pulmonary insufficiency, cardiomyopathy
<i>LCK</i> deficiency	Retinal vasculitis	Pericarditis
DNA repair defects		
Ataxia–telangiectasia	Telangiectasias, ischemic heart disease	Mitral valve prolapse, mitral/tricuspid insufficiency
Nijmegen breakage syndrome		VSD, PDA
Predominantly antibody deficiencies		
CVID	Aortic root dilatation, thoracic and abdominal aortic aneurysms, abnormal aortic arch	Giant-cell myocarditis
TWEAK deficiency		Cardiomyopathy
Defects of phagocyte numbers, function, or both		
Severe congenital neutropenia type 4/ <i>G6PC3</i> deficiency	Anomalous pulmonary veins	ASD, PDA, mitral valve anomalies, pulmonary stenosis Cor triatriatum
Barth syndrome	Thromboembolic cerebrovascular events	Dilated/hypertrophic cardiomyopathy Left ventricle non-compaction Ventricular arrhythmias Endocardial fibroelastosis
Cohen syndrome		Decreased LV function Mitral valve prolapse
Shwachman-Diamond syndrome		Dilated cardiomyopathy VSD ASD PDA
Warts Hypogammaglobulinemia Infections and Myelokathexis (WHIM syndrome)		Tetralogy of Fallot
MonoMAC syndrome (<i>GATA2</i> deficiency)	Capillary leak syndrome with lymphedema	
Chronic granulomatous disease	Abnormal vascular tone Coronary arterial disease Cerebral aneurysms	Aortic/mitral insufficiency
Well-Defined Syndromes with Immunodeficiency		
DiGeorge syndrome	Aortic arch defects Coarctation of aorta Abnormal subclavian artery	Conotruncal abnormalities: Tetralogy of Fallot Truncus arteriosus Interrupted aortic arch TGA ASD VSD Pulmonary stenosis

Continues

Appendix A1: (Continued)

PID	Vascular abnormalities	Cardiac abnormalities
CHARGE syndrome	Aortic arch defects	Conotruncal abnormalities: Tetralogy of Fallot AVSD ASD VSD PDA
Wiskott-Aldrich syndrome	Aortic/systemic aneurysms Aortic root dilatation Small/medium/large vessel vasculitis	
Roifman Syndrome		Left ventricle Non-compaction
Schimke immune-osseous dysplasia	Moya-moya vasculitis Cerebral arteries stenosis Diffuse aortic narrowing	
Hyper IgE Syndromes: AD HIES (STAT3 deficiency) AR HIES (DOCK8 deficiency)	Systemic vasculitis Coronary/thoracic/aortic aneurysms Moya-moya vasculitis Brain infarctions Aortic aneurysms	
VODI syndrome	Hepatic veno-occlusive disease	Left atrial endocardial fibrosis
ICF syndrome		VSD ASD
FILS syndrome	Telangiectasias Livedo reticularis	
Disease of Immune Dysregulation		
X-linked lymphoproliferative disorder type 1	Systemic small/medium vessel vasculitis: coronary, cerebral, retinal	
FADD deficiency		Pulmonary atresia with VSD, Left sided superior vena cava draining into left atrium
Aicardi-Goutières syndrome	Vasculopathy of brain and retina Stenosis of carotid Intracranial vessel stenosis Small vessel bleeding	
Autoinflammatory Disorders		
HOIL1 Deficiency		Dilated cardiomyopathy

Note: Acronyms are defined in Appendix B1.

Appendix A2: Primary immunodeficiency disorders associated with cardiac abnormalities.

Cardiac abnormalities	PID
Atrial septal defect	STK4 deficiency SCN4/G6PC3 deficiency DiGeorge syndrome CHARGE syndrome Shwachman-Diamond syndrome ICF syndrome
Ventricular septal defect	Nijmegen breakage syndrome DiGeorge syndrome CHARGE syndrome Shwachman-Diamond syndrome ICF syndrome ALPS-FADD deficiency
Patent ductus arteriosus	Nijmegen breakage syndrome SCN4/G6PC3 deficiency DiGeorge syndrome CHARGE syndrome Shwachman-Diamond syndrome
Pulmonary stenosis/atresia	SCN4/G6PC3 deficiency DiGeorge syndrome ALPS-FADD deficiency
Mitral valve prolapse	Ataxia-telangiectasia Cohen Syndrome
Valve insufficiency	STK4 deficiency: mitral/tricuspid/pulmonary Ataxia telangiectasia: mitral/tricuspid SCN4/G6PC3 deficiency: mitral CGD: aortic/mitral
Conotruncal anomalies	DiGeorge syndrome CHARGE syndrome
Tetralogy of Fallot	WHIM syndrome DiGeorge syndrome CHARGE syndrome
Aortic arch abnormalities	CVID DiGeorge syndrome CHARGE syndrome
Ventricular noncompaction	Roifman syndrome Barth syndrome
Cardiomyopathy	Omenn syndrome ORAI1 deficiency STK4 deficiency TWEAK deficiency Barth syndrome (dilated/hypertrophic) Cohen syndrome Shwachman-Diamond syndrome HOIL1 Deficiency
Myocarditis	CVID
Pericarditis	LCK deficiency
Arrhythmias	ORAI1 deficiency Barth syndrome

Note: Acronyms are defined in Appendix B1.

Appendix A3: Primary immunodeficiency disorders associated with vascular abnormalities.

Vascular abnormalities	PID
Aortic root dilatation	CVID Wiskott-Aldrich syndrome
Aortic Aneurysms	CVID Wiskott-Aldrich syndrome AD and AR Hyper IgE syndromes
Coronary aneurysms	AD Hyper IgE syndrome
Intracranial aneurysms	AD chronic mucocutaneous candidiasis Chronic granulomatous disease
Cerebral vasculitis	AD chronic mucocutaneous candidiasis Wiskott-Aldrich syndrome Schimke immune-osseous dysplasia X-linked lymphoproliferative disorder type 1
Moya-Moya vasculitis	AR Hyper IgE syndrome Schimke immune-osseous dysplasia
Coronary vasculitis	X-linked lymphoproliferative disorder type 1
Retinal vasculitis	LCK deficiency X-linked lymphoproliferative disorder type 1
Systemic vasculitis	Wiskott-Aldrich syndrome X-linked lymphoproliferative disorder type 1
Systemic vessel stenosis	Aicardi-Goutières syndrome: brain/carotid/retina
Abnormal vascular tone	Chronic granulomatous disease
Aortic arch defects	DiGeorge syndrome CHARGE syndrome
Abnormal pulmonary veins	SCN4/G6PC3 deficiency
Ischemic heart disease	Ataxia-telangiectasia Chronic granulomatous disease
Thromboembolic cerebrovascular events	Barth syndrome
Telangiectasias	Ataxia-telangiectasia FILS syndrome
Capillary leak syndrome	LCK deficiency MonoMAC (GATA2 deficiency)
Veno-occlusive disease of the liver	VODI syndrome

Note: Acronyms are defined in Appendix B1.

Appendix B1: Acronyms.

AGS: Aicardi–Goutières syndrome
ALPS: Autoimmune lymphoproliferative syndrome
AT: Ataxia–telangiectasia
CGD: Chronic granulomatous disease
CHARGE: Coloboma, Heart, Atresia choanae, Retardation, Genital and Ear abnormalities
CMCC: Chronic mucocutaneous candidiasis
CVID: Common variable immunodeficiency
CVS: Cardiovascular system
DGS: DiGeorge Syndrome
FILS: Facial dysmorphism, Immunodeficiency, Livedo, and Short stature syndrome
G6PC3: Glucose-6-phosphatase, catalytic subunit 3
HIES: Hyper-IgE Syndrome
HSCT: Hematopoietic stem cells transplantation
ICF: Immunodeficiency with centromeric instability and facial anomalies
PID: Primary immunodeficiency diseases
SCID: Severe combined immunodeficiency
SIOD: Schimke immuno-osseous dysplasia
SCN: Severe congenital neutropenia
SDS: Shwachman–Diamond syndrome
TREC: T cell receptor excision circles
TWEAK: TNF-like weak inducer of apoptosis
VODI: Veno-occlusive disease with immunodeficiency
WAS: Wiskott–Aldrich Syndrome
WHIM: Warts, Hypogammaglobulinemia, Infections, and Myelokathexis
XLP: X-linked lymphoproliferative disease